

RESEARCH ARTICLE

Open Access

# A systematic review of the overlap of fluid biomarkers in delirium and advanced cancer-related syndromes



Ingrid Amgarth-Duff<sup>1\*</sup>, Annmarie Hosie<sup>1</sup>, Gideon Caplan<sup>2,3</sup> and Meera Agar<sup>1,4,5</sup>

## Abstract

**Background:** Delirium is a serious and distressing neurocognitive disorder of physiological aetiology that is common in advanced cancer. Understanding of delirium pathophysiology is largely hypothetical, with some evidence for involvement of inflammatory systems, neurotransmitter alterations and glucose metabolism. To date, there has been limited empirical consideration of the distinction between delirium pathophysiology and that of the underlying disease, for example, cancer where these mechanisms are also common in advanced cancer syndromes such as pain and fatigue. This systematic review explores biomarker overlap in delirium, specific advanced cancer-related syndromes and prediction of cancer prognosis.

**Methods:** A systematic review (PROSPERO CRD42017068662) was conducted, using MEDLINE, PubMed, Embase, CINAHL, CENTRAL and Web of Science, to identify body fluid biomarkers in delirium, cancer prognosis and advanced cancer-related syndromes of interest. Studies were excluded if they reported delirium tremens only; did not measure delirium using a validated tool; the sample had less than 75% of participants with advanced cancer; measured tissue, genetic or animal biomarkers, or were conducted post-mortem. Articles were screened for inclusion independently by two authors, and data extraction and an in-depth quality assessment conducted by one author, and checked by two others.

**Results:** The 151 included studies were conducted in diverse settings in 32 countries between 1985 and 2017, involving 28130 participants with a mean age of 69.3 years. Seventy-one studies investigated delirium biomarkers, and 80 studies investigated biomarkers of an advanced cancer-related syndrome or cancer prognosis. Overall, 41 biomarkers were studied in relation to both delirium and either an advanced cancer-related syndrome or prognosis; and of these, 24 biomarkers were positively associated with either delirium or advanced cancer syndromes/prognosis in at least one study. The quality assessment showed large inconsistency in reporting.

**Conclusion:** There is considerable overlap in the biomarkers in delirium and advanced cancer-related syndromes. Improving the design of delirium biomarker studies and considering appropriate comparator/controls will help to better understanding the discrete pathophysiology of delirium in the context of co-existing illness.

**Keywords:** Delirium, Biomarker, Advanced cancer, Review

\* Correspondence: [Ingrid.Amgarth-Duff@uts.edu.au](mailto:Ingrid.Amgarth-Duff@uts.edu.au)

<sup>1</sup>University of Technology Sydney, Faculty of Health, IMPACCT -Improving Palliative, Aged and Chronic Care through Clinical Research and Translation, Sydney, NSW, Australia

Full list of author information is available at the end of the article



© The Author(s). 2020 **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>. The Creative Commons Public Domain Dedication waiver (<http://creativecommons.org/publicdomain/zero/1.0/>) applies to the data made available in this article, unless otherwise stated in a credit line to the data.

## Background

Delirium is a very common cause of acute cognitive change in people with advanced cancer [1] and is associated with increased morbidity and mortality [2, 3]. Delirium is a serious and complex neurocognitive disorder characterized by acute deterioration in attention, awareness and cognition, variously affecting memory, language and visuospatial ability, orientation and perception [4].

Delirium occurs in people who are medically unwell, due to the underlying disease which has put them at risk (e.g. dementia, cancer, infection, renal impairment) or intercurrent problems, and the subsequent medical treatment (e.g. surgery, medication). Delirium can occur for any person, with those who are older, have advanced illness, and/or prior cognitive impairment most at risk [5]. The prevalence of delirium in patients with advanced cancer in oncology and palliative care settings is higher than that in most other settings, including geriatrics [1, 6–9]. A systematic review of palliative care patients (with 98.9% of participants with advanced cancer), reported delirium incidence rates between 3% and 45%. Delirium prevalence ranged from 13.3% to 42.3% at admission to hospital, and 25% to 62% during admission. Delirium prevalence increased up to 88% in the hours to days before death [1].

The pathophysiology of delirium is poorly understood, and largely hypothetical. Current hypotheses include: neuronal ageing, neuroinflammation, oxidative stress, neuroendocrine dysregulation, disruption to the circadian rhythm, and neurotransmitter dysregulation [10, 11]. A reduction in glucose metabolism seen in people with delirium is a model with developing evidence [12, 13]. Collectively, the biological correlates of delirium are referred to as 'delirium biomarkers'. A biomarker is a biological molecule found in blood, other body fluids, or tissues that is a sign of a normal or abnormal process, or of a condition or disease [14]. Biomarkers are most commonly studied to investigate their correlation with a disease in order to better understand its underlying pathophysiology, and subsequently inform prevention and treatment strategies for that disease. A challenge for the field of delirium research is that correlation may exist between biomarkers of delirium and those of the patient's disease or injury which placed them at increased risk of delirium, or which precipitated it (for example sepsis or hip fracture). Such correlation should be factored into delirium biomarker research, yet rarely has been. Better understanding of the interplay between delirium pathophysiology and that of correlated conditions and diseases, for example, cancer (the focus of this review), is crucial to develop more effective prevention and treatment of delirium.

We therefore conducted a systematic review of the literature to explore the overlap between biomarkers that

have been studied in delirium and biomarkers that have been studied in cancer-related syndromes. Our aim was to identify biomarkers associated with delirium and with specific clinical situations in advanced cancer (namely prognosis; cognitive impairment, anorexia cachexia, cancer pain, cancer-related fatigue, and sickness behavior); and to evaluate the nature and extent of overlap of the findings.

## Methods

A systematic review according to the Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA) [15] was conducted. In July 2017, two separate searches were conducted in MEDLINE, PubMed, Embase, CINAHL, CENTRAL, and Web of Science. The first was for literature of delirium biomarkers; the second was for literature of biomarkers in advanced cancer-related syndromes. Primary terms for the delirium search were: 'delirium' and 'biomarker'. Search terms for the cancer search were: 'cancer', 'neoplasms', 'metastasis', 'fatigue', 'sickness behavior', 'cancer pain', 'cachexia', and 'prognosis'. Additional terms which encompassed commonly researched biomarkers were also included. Filters in Medline were: 1: Humans; 2: English language and 3: Published from 1980 onward (when delirium was first included in the *DSM, Third Edition (DSM-III)*). Search terms and filters were tailored to each subsequent database, as required. The full search strategy is provided in Additional file 1. Reference lists of included studies and relevant systematic reviews and meta-analyses identified in the search were examined for additional eligible studies.

We included English language studies published in peer-reviewed journals that reported body fluid biomarkers in adult participants with delirium, cancer prognosis or an advanced cancer-related syndrome of interest. Studies were excluded if they reported delirium tremens only; did not measure delirium using a validated tool; the sample had less than 75% of participants with advanced cancer; measured tissue, genetic or animal biomarkers, or were conducted post-mortem. Protocols and ongoing studies were also excluded. Based on the expert knowledge of the authors in both delirium and cancer, the advanced cancer-related syndromes and prognosis were chosen based on the potential biological plausibility that the pathophysiological mechanisms could overlap with that of delirium. We limited the search to advanced cancer as this is the cancer population with the highest prevalence of both delirium and the cancer-related syndromes of interest.

The following definitions were used in this review:

**Anorexia cachexia:** A complex metabolic syndrome of involuntary weight loss associated with cancer and some other palliative conditions [16].

**Cancer related fatigue:** A distressing, persistent, subjective sense of physical, emotional, and/or cognitive tiredness or exhaustion related to cancer and/or cancer treatment that is not proportional to recent activity and interferes with usual functioning [17].

**Cancer-related pain:** An unpleasant sensory and emotional experience associated with actual or potential tissue damage, or described in terms of such damage [18].

**Cancer-related cognitive impairment:** Cognitive impairment that is commonly experienced by cancer patients and those in remission. The cognitive domains most commonly affected are memory, concentration, information processing speed and executive function [19].

**Sickness behaviour:** The coordinated set of behavioural changes that develop in sick individuals during the course of an infection. Sickness behavior is also seen in other illness including cancer [20, 21].

**Cancer prognosis:** The likely outcome or course of the disease; the chance of recovery or recurrence. Cancer prognosis is assessed by cancer-specific survival, overall survival, progression free survival or relative survival [22].

Search results were imported into Endnote X7 software, duplicates removed and then exported into Covidence™ ([www.covidence.org](http://www.covidence.org)). Two reviewers per search (IAD and AH: delirium search, IAD and MA: cancer search) independently applied eligibility criteria for both searches and examined title and abstracts. Exclusions were documented only for articles that required full-text to make a formal decision. Inter-reviewer disagreement on included studies was discussed to resolve any discrepancies, with the third reviewer consulted when required. Data extraction was conducted by one reviewer (IAD) using Excel (2016) with two other reviewers (MA and AH) providing input and oversight. Data extraction was guided by the REporting recommendations for tumor MARKer prognostic studies (REMARK) checklist [23].

In the absence of a gold standard risk of bias assessment for biomarker studies, one reviewer (IAD) applied an adaptation of the REMARK checklist [23] to assess the methodological quality of the included studies, with 10% verification by two other reviewers (MA and AH).

The heterogeneity of data precluded performing a meta-analysis; we therefore reported the data using a narrative synthesis approach with text and tabular summaries. The synthesis was structured according to the overlap of the biomarkers in delirium, cancer prognosis and the cancer syndromes, the biomarker type, assay used, and numbers and proportions of participants who had delirium and advanced cancer. We defined 'overlap' as any biomarker that was studied in both a delirium study and an advanced cancer syndrome study.

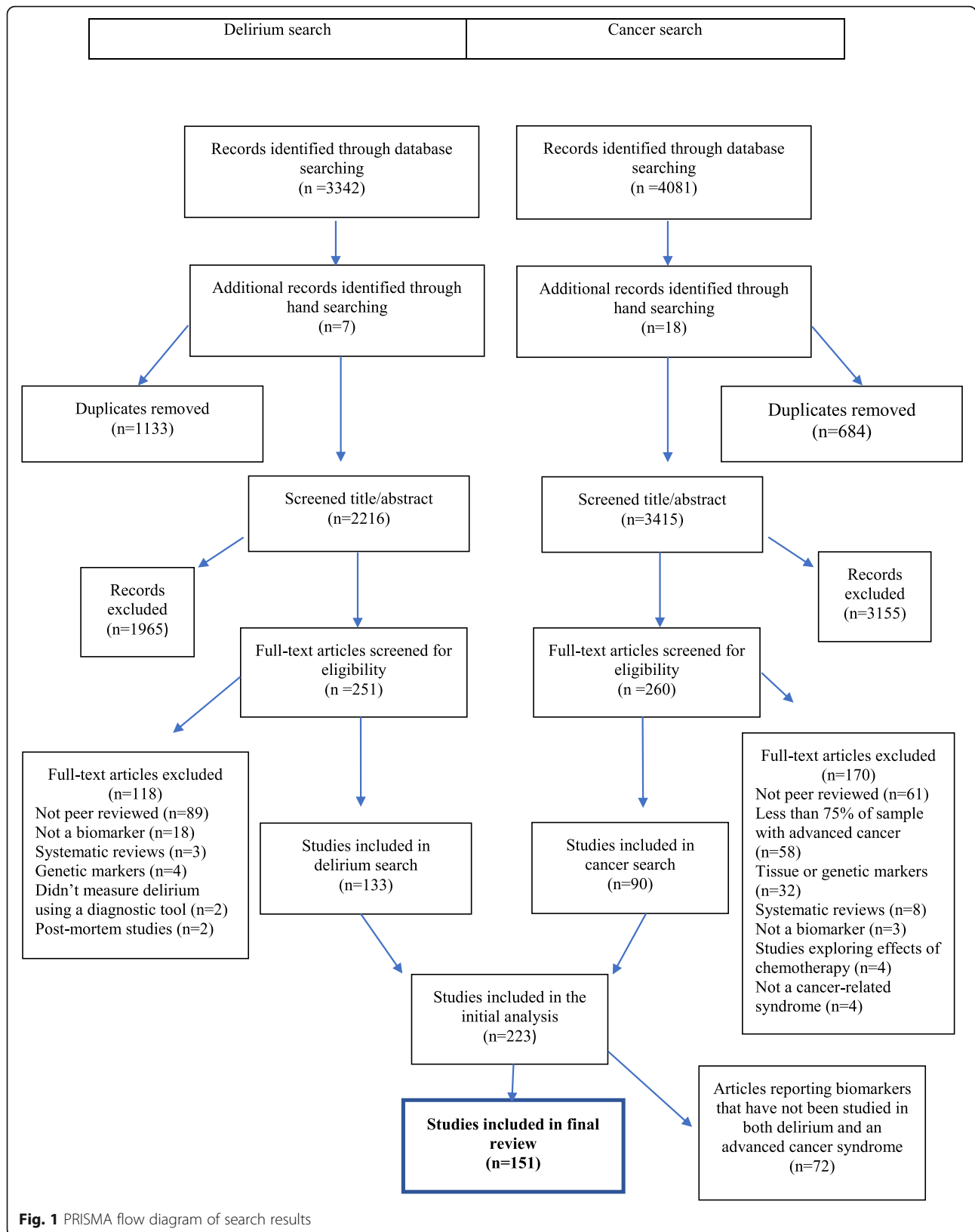
## Results

The delirium search yielded 3342 articles and the cancer syndromes search 4081, giving a total of 7423 articles. An additional 25 articles were found through the hand search. After removal of 1817 duplicates and 5120 articles through title and abstract screening, we reviewed 511 full text papers and subsequently excluded 288. After initial analysis, a further 72 were excluded as they did not report a biomarker studied in delirium and advanced cancer. This resulted in a total of 151 articles included in this review: 71 reported biomarkers studied in delirium, and 80 reported biomarkers studied in a cancer syndrome or prognosis (Figure 1).

The 151 studies were conducted between 1985 and 2017 in Europe ( $n=86$ ), Asia ( $n=33$ ), The Americas ( $n=27$ ), Australia ( $n=2$ ), and multiple regions ( $n=3$ ). Studies were set in a large range of settings, with the most common in general hospital settings ( $n=111$ ; 73%). Thirty-nine studies (26%) did not report the setting. Sample sizes ranged from 7-2456, with relatively even numbers of male and female participants (55.4% male). Ninety-nine articles reported a mean age, with an overall weighted mean age of 69.3 years. Of the 37 articles that reported the median age of participants, the overall median age was 54.5 years. The overall age of participants in the remaining 15 articles was not possible to determine (Additional files 2 and 3). Blood biomarkers were examined in 138 studies, 4 studies examined biomarkers in cerebrospinal fluid (CSF), 3 in urine, and 16 (11%) did not report the type of biological material. Of the studies that reported the assay technique, diverse assays were used ( $n=20$ ), with Enzyme-linked immunosorbent assay (ELISA) being the most common ( $n=62$ ; 58%). Forty-four studies (29%) did not report the specific assay used. Of these, 21 studies (48%) were routinely measured biomarkers (Tables 1 and 2).

A total of 41 biomarkers were found to be common in both delirium and advanced cancer syndrome studies. The five most commonly studied biomarkers were C-reactive protein (CRP) ( $n=79$ ), interleukin (IL)-6 ( $n=58$ ), tumor necrosis factor alpha (TNF- $\alpha$ ) ( $n=42$ ) IL-10 ( $n=21$ ) and IL-8 ( $n=24$ ). Of these, 24 biomarkers had a positive association with delirium, cancer prognosis or a cancer syndrome in at least one study. No cancer studies reported having any participants with delirium, and of the delirium studies, six reported participants with cancer. Figure 2 illustrates two main populations identified from this systematic review, with the centre showing the 'true overlap' defined as studies that included participants with both delirium and cancer ( $n=6$  studies).

In two of these studies, all participants in the study had cancer; in another, 64.2% of participants had cancer; in the remaining three studies, less than 30% of all participants had cancer. In three of the studies, 100% of



**Table 1** Characteristics of assays and main findings of included delirium studies\*

Author and year	Participants		Endpoints	Biomarkers studied	Biological material	Assay method	Covariates accounted for in multivariate analysis	Results	Negative association with at least one delirium endpoint **
	Total (N)	Sample							
Egberts et al. (2017) [24]	86	Aged ≥65 admitted to geriatrics	Delirium presence	CRP, NLR	Blood	Flow cytometry	Age, gender, the CCI score, CRP level, and WBC counts	NLR	CRP
Kozak et al. (2017) [25]	60	Patients with acute ischemic stroke	Delirium presence	TNF-α, IL-1β, IL-18, BDNF, NSE	Serum	ELISA	No multivariate analysis	None	TNF-α, IL-1β, IL-18, BDNF, NSE
Tomasi et al. (2017) [26]	38	Patients with sepsis-associated delirium and non-sepsis associated delirium <sup>a</sup>	Delirium presence	IL-6, IL-8, IL-10, BDNF, VCAM-1, ICAM-1, MPO, cathepsin, PDGF-AA, PDGF-AB/BB, RANTES, PAI, NCAM	Plasma	ELISA	No multivariate analysis	IL-6, IL-10, RANTES, VCAM-1, ICAM-1, PDGF-AB/BB	IL-8, MPO, BDNF, NCAM, PDGF-AA, PAI, Cathepsin D
Vasulilashorn et al. (2017) [27]	560	Patients ≥70 undergoing major non-cardiac surgery <sup>a</sup>	-Delirium incidence -Delirium duration -Delirium severity	CRP	Plasma	ELISA	Age, sex, surgical procedure, anesthesia route, CCI and POST-OP infectious complications	CRP	None
Chu et al. (2016) [28]	103	Patients aged ≥70 admitted for acute or elective vertebral, knee, or hip surgery	Delirium incidence	IGF-1	Serum	ELISA	MMSE and age	None	IGF-1
Dillon et al. (2016) [28]	Entire sample (n=566); pooled sample non-cardiac surgery <sup>a</sup> (n=150)	Advanced cancer excluded; other cancer stages NR	Delirium incidence	Proteomics <sup>b</sup>	Plasma	ELISA	No multivariate analysis	CRP (PRE-OP, PACU, POD2)	CRP (PO1MO)
Guo et al. (2016) [29]	572	Aged ≥65 with hip fractures undergoing THA <sup>a</sup>	-Delirium presence -Delirium prevalence	CRP, Alb, Hb	Blood	NR	NR	CRP, Alb, Hb	None
Karlicic et al. (2016) [30]	120	Patients with delirium in the psychiatric ICU	Lethal outcome	CRP	NR	NR	Age, pneumonia and CRP	CRP	None
Neerland et al. (2016) [31]	149	Patients with acute hip fracture	Delirium presence	CRP, IL-6, sIL-6R	CSF	ELISA	No multivariate analysis	CRP <sup>b</sup>	sIL-6R, IL-6
Shen et al. (2016) [32]	140	Patients ≥65 undergoing elective gastrointestinal tumor resection <sup>a</sup>	-Delirium incidence -Delirium severity	IGF-1, CRP, IL-6	Serum	ELISA	NR	IGF-1, CRP, IL-6	None
Sun et al. (2016) [33]	112	Oral cancer patients <sup>a</sup>	Delirium incidence	IL-6, CRP, PCT, cortisol, AB1-40	Blood	ELISA	No multivariate analysis	IL-6, CRP, PCT, cortisol, AB1-40	None
Yen et al. (2016) [34]	98	Patients undergoing elective knee replacement surgery	Delirium incidence	IGF-1	Serum	ELISA	Obstructive sleep apnea, IGF-1 and diabetes	None	IGF-1
Avila-Funes et al. (2015) [35]	141	Patients aged ≥70 admitted to tertiary care hospital	Delirium incidence	Cortisol, E2	Blood	Radioimmunoassay	Age, BMI, comorbidity, MMSE, previous history of delirium, BUN/C ratio, and cortisol levels	E2	Cortisol
Brum et al. (2015) [36]	70	Oncology inpatients <sup>a</sup>	Delirium presence	BDNF, TNF-α	Serum	ELISA + Flow cytometry	No multivariate analysis	None	BDNF, TNF-α
Egberts et al. (2015) [37]	86	Patients admitted	Delirium	NP, IL-6, IGF-1	Plasma	HPLC	Age, gender and the CCI	NP, IL-6, IGF-1	None

**Table 1** Characteristics of assays and main findings of included delirium studies\* (Continued)

Author and year	Participants Total (N)	Sample	Total participants with cancer/total participants in the study	Number of delirium with cancer/total number delirium (%)	Endpoints	Biomarkers studied	Biological material	Assay method	Covariates accounted for in multivariate analysis	Results	Negative association
[2015] [37]		To Internal Medicine and Geriatrics <sup>a</sup>		measured/ NR	presence					Positive association with at least one delirium endpoint **	
<b>Foroughan et al.</b> (2015) [38]	200	Elderly patients admitted to general hospital	18/200 (9)	12/44 (27)	Delirium presence	CRP, Hb	Blood	NR	and those including NP were adjusted for age, gender, CCI, tertiles of eGFR and CRP	CRP, Hb	None
Skrede et al. (2015) [39]	10	Patients with hip fracture	Not measured/NR	Not measured/ NR	Delirium incidence	MCP-1	Serum	ELISA	No multivariate analysis	MCP-1	None
Vasurilashorn et al. (2015) [40]	566	Patients ≥70 undergoing major non-cardiac surgery <sup>a</sup>	Not measured/NR	Not measured/ NR	Delirium incidence	IL-18, IL-2, IL-4, IL-5, IL-6, IL-8, IL-10, IL-12, IFN-γ, GM-CSF, TNF-α, VEGF	Plasma	Luminex assay	No multivariate analysis	IL-18, IL-2, IL-6, IL-8, IL-12, VEGF, IL-5, TNF-α	GM-CSF, IFN-γ, IL-10, IL-4
Alexander et al. (2014) [41]	77	ICU patients requiring mechanical ventilation	Not measured/NR	Not measured/ NR	-Delirium presence -Delirium duration	IL-6, IL-8, IL-10, APOE	Serum	ELISA	Age, sex, APACHE III, CCI, 24-hour propofol dose, 24-hour narcotic dose, and 24-hour benzodiazepine dose.	APOE	IL-10, IL-8, IL-6
Baranyi et al. (2014) [42]	34	Patients undergoing surgery for CPB <sup>a</sup>	Not measured/NR	Not measured/ NR	Delirium incidence	sIL-2R	Serum	ELISA	No multivariate analysis	sIL-2R	None
Cape et al. (2014) [43]	43	Patients >60 years old with hip fracture	Not measured/NR	Not measured/ NR	-Delirium incidence -Delirium prevalence	IL-1β, IFN-γ, GFAP, IGF-1, IL-1RA	CSF	ELISA	Presence of prior dementia	IL-1β, IL-1RA <sup>c</sup>	GFAP, IFN-γ, IGF-1
Capri et al. (2014) [44]	351	Patients admitted for any kind of emergency or elective surgery <sup>a</sup>	Comorbidity measured, cancer NR	Comorbidity measured, cancer NR	Delirium presence	IL-1β, IL-2, IL-6, IL-8, IL-10, TNF-α	Plasma	ELISA	Age, comorbidity, ADL, IADL, HADS, and pre-op benzodiazepines intake	IL-6, IL-2	IL-8, IL-10, IL-1β (UDL), TNF-α (UDL)
Chen et al. (2014) [45]	372	Patients aged 265 who underwent surgery for a femoral neck fracture or an intertrochanteric fracture <sup>a</sup>	Not measured/NR	Not measured/ NR	Delirium presence	LP	Plasma	ELISA	Age, ASA, type of surgery and plasma leptin level	LP	None
Hatta et al. (2014) [46]	29	Patients aged 65-89 admitted to hospital due to an emergency	Not measured/NR	Not measured/ NR	Delirium incidence	NK cell activity, IL-1β	Blood	ELISA	No multivariate analysis	NK cell activity	IL-1β
Kazmierski et al. (2014) [47]	113	ICU patients scheduled for CABG surgery with CPB	Not measured/NR	Not measured/ NR	Delirium incidence	Cortisol, IL-2, TNF-α, HCY, cobalamin	Serum	CLIA	NR	Cortisol, IL-2, TNF-α, HCY	Cobalamin
Pitche et al. (2014) [48]	710	Patients admitted to a Medical Acute Admission Unit	Not measured/NR	Not measured/ NR	-Delirium incidence severity	CRP	NR	NR	NR	CRP	None
Ritter et al. (2014) [49]	78	ICU patients	Not measured/NR	Not measured/ NR	Delirium presence	TNF-α, STNFR-1, STNFR2, APN, IL-1β, IL-6, IL-10	Plasma	ELISA	Sedation and sepsis	STNFR-1, STNFR2, IL-1β	TNF-α, IL-6, IL-10
Zhang et al. (2014) [50]	223	ICU patients	Not measured/NR	Not measured/ NR	Delirium presence	CRP	Plasma	i-CHROMATM	Age, sex, APACHE II, intubation status, living alone, physical restraint, alcohol drinking, smoking, type of medical condition, and hospital LOS before ICU admission	CRP	None

**Table 1** Characteristics of assays and main findings of included delirium studies\* (Continued)

Author and year	Participants Total (N)	Sample	Total participants with cancer/total participants in the study	Number of delirium with cancer/total number delirium (%)	Endpoints	Biomarkers studied	Biological material	Assay method	Covariates accounted for in multivariate analysis	Results	Negative association with at least one delirium endpoint **
Cerejeira et al. (2013) [51]	101	Patients ≥60 years without dementia undergoing elective hip arthroplasty <sup>a</sup>	Not measured/NR	Not measured/NR	Delirium incidence	Cortisol, IGF-1, CRP, IL-6, IL-8, IL-10	Plasma	ELISA	No multivariate analysis	Cortisol	CRP, IL-6, IL-8, IL-10, IGF-1
Colkesen et al. (2013) [52]	52	Patients with ACS admitted to coronary ICU <sup>b</sup>	Not measured/NR	Not measured/NR	Delirium presence	Cortisol, troponin I, MB-CK	Serum	CLIA	NR	Cortisol	Troponin I, MB-CK
Kazmierski et al. (2013) a [53]	113	ICU patients scheduled for CABG surgery with CPB	Not measured/NR	Not measured/NR	Delirium incidence	Cortisol, IL-2	Plasma	CLIA	NR	Cortisol <sup>d</sup> , IL-2	None
Kazmierski et al. (2013) b [54]	113	ICU patients scheduled for CABG surgery with CPB	Not measured/NR	Not measured/NR	Delirium incidence	IL-2, TNF-α	Plasma	CLIA	NR	IL-2, TNF-α	None
Liu et al. (2013) [55]	338	Patients aged ≥60 undergoing major non-cardiac surgery <sup>a</sup>	Not measured/NR	Not measured/NR	Delirium incidence	IL-6	Blood	ELISA	Age, education, history of coronary artery disease, alcohol, history of alcoholism, PRE-OP ASA ≥ 3, PRE-OP NYHA ≥ 2, PRE-OP MMSE score ≤ 24, PRE-OP serum IL-6 ≥ 7.5 pg/ml, POST-OP serum IL-6, POST-OP VAS pain level	IL-6	None
Plaschke et al. (2013) [56]	114	1. Patients following heart surgery <sup>a</sup> 2. Patients on the non-cardiac ICU <sup>a</sup>	Not measured/NR	Not measured/NR	Delirium incidence	IL-6	Plasma	ELISA	No multivariate analysis	None	IL-6
Skrobik et al. (2013) [57]	99	ICU patients <sup>a</sup>	Not measured/NR	Not measured/NR	Drug-induced coma and delirium	TNF-α, IL-1β, IL-1RA, IL-6, IL-8, IL-10, IL-17, MIP-1β, MCP-1	Blood	BCA	Fentanyl, midazolam, CYP3A4/5, P-gp inhibitors	IL-6	TNF-α, IL-17, IL-8, MCP-1, IL-1RA, MIP-1β, IL-10, IL-1β
Westhoff et al. (2013) [58]	61	Patients ≥75 admitted for surgical repair of acute hip fracture <sup>b</sup>	Not measured/NR	Not measured/NR	Delirium incidence	EGF, eotaxin, FGF-2, Flt-3L, Fractalkine, G-CSF, GM-CSF, IFN-α2, IFN-γ, IL-1, IL-1RA, IL-1α, IL-1β, IL-2, sIL-2Rα, IL-3, IL-4, IL-5, IL-6, IL-7, IL-8, IL-9, IL-10, IL-12p40, IL-12p70, IL-13, IL-15, IL-17, IP-10, MCP-1, MCP-3, MDC, MIP-1α, MIP-1β, PDGF-AA, PDGF-AB/BB, RANTES, sCD40L, TGF-α, TNF-α, TNF-β, VEGF	Blood + CSF	Lumbar punctures and Luminex assays	No multivariate analysis	Flt-3L, IL-1RA, IL-6	EGF, eotaxin, FGF-2, Fractalkine, G-CSF, GM-CSF, IFN-α2, IFN-γ, IL-1, IL-1β, IL-2, sIL-2Rα, IL-3, IL-4, IL-5, IL-6, IL-7, IL-8, IL-9, IL-10, IL-12p40, IL-12p70, IL-13, IL-15, IL-17, IP-10, MCP-1, MCP-3, MDC, MIP-1α, MIP-1β, PDGF-AA, PDGF-AB/BB, RANTES, sCD40L, TGF-α, TNF-α, TNF-β, VEGF
Bakker et al. (2012) [59]	201	Patients undergoing cardiac surgery	Not measured/NR	Not measured/NR	Delirium incidence	Cre	Plasma	NR	NR	Cre	None
Baranyi et al. (2012) [60]	34	Patients undergoing surgery for cardiopulmonary bypass <sup>a</sup>	Not measured/NR	Not measured/NR	Delirium incidence	Alb, CRP	Serum	NR	No multivariate analysis	Alb	CRP
Cerejeira et al. (2012) [61]	101	Patients aged ≥60 undergoing elective total hip arthroplasty <sup>a</sup>	Not measured/NR	Not measured/NR	Delirium incidence	IL-8, IL-1β, IL-6, IL-10, TNF-α, CRP, AChE, BuChE	Blood	ELISA (Multiplex assay)	No multivariate analysis	AChE, BuChE	CRP, IL-1β, TNF-α, IL-6, IL-10
Girard et al. (2012) [62]	138	Mechanically ventilated ICU patients <sup>a</sup>	Not measured/NR	Not measured/NR	Delirium incidence	CRP, MMP-9, MPO, NGAL, sTNFR1, D-dimer, protein C, PAI-1, WVF	Plasma	ELISA	Age, severity of illness, and severe sepsis	MMP-9, Protein C, sTNF-R1	CRP, MPO, NGAL, D-dimer, PAI-1, WVF
Oxse et al. (2012) [63]	125	Patients ≥70 undergoing elective cardiac	Not measured/NR	Not measured/NR	Delirium incidence	NP, BH4, HVA, Gly, Ser, Gly, Cit, Tau, Arg, Met, Tyr, Phe, Leu, Ile, Val, Ile, Val, Trp, LNAa, TyrlNAa, PheLNAa	Plasma	HPLC	BH4, total biotin, HVA, ratios of TrpLNAa, TyrLNAa, pheLNAa, phe: LNAa, phe: Tyr, Cit:Arg,	NP, HVA	BH4, Gly, Ser, Gly, Cit, Tau, Arg, Met, Tyr, Phe, Leu, Ile, Val, TrpLNAa, TyrlNAa, PheLNAa,

**Table 1** Characteristics of assays and main findings of included delirium studies\* (Continued)

Author and year	Participants Total (N)	Sample	Total participants with cancer/total participants in the study	Number of delirium with cancer/total number delirium (%)	Endpoints	Biomarkers studied	Biological material	Assay method	Covariates accounted for in multivariate analysis	Results	Positive association with at least one delirium endpoint **	Negative association	
		surgery											
						LNAa, Phetyr, Citarg, TauSer 9 met						Phetyr, Citarg, TauSer 9 met	
Bisschop et al. (2011) [64]	143	Patients undergoing surgery for hip fracture	Not measured/NR	Not measured/NR	-Delirium presence -Delirium severity	Cortisol, insulin, glucose	Blood	NR	TSM ratio; baseline CRP, plasma urea, cre, age, sex, type of surgery, acute cardiac surgical risk factors, EuroSCORE, MMSE, pre-op anxiety and depression, and chronic medical comorbidity	Cortisol		Glucose, insulin	
Holmes et al. (2011) [65]	222	Patients with mild to severe AD	Not measured/NR	Not measured/NR	-Presence of sickness behaviour -Delirium incidence	IL-6, TNF-α, CRP	Blood	ELISA	Baseline ADAS score, age, gender, and the presence of delirium	None		IL-6, TNF-α, CRP	
Lee et al. (2011) [66]	65	Patients 265 who had undergone hip surgery	Not measured/NR	Not measured/NR	Delirium incidence	CRP	Blood	NR	No multivariate analysis	None		CRP	
McGrane et al. (2011) [67]	87	Mechanically ventilated, medical and surgical ICU patients <sup>a</sup>	Not measured/NR	Not measured/NR	Delirium/ coma-free days	PCT, CRP	Blood	TRACE Assay analysis	Age, APACHE II, sedation group (dexmedetomidine vs. lorazepam), and sepsis	PCT		CRP	
Morandi et al. (2011) [68]	110 <sup>e</sup>	Mechanically ventilated medical ICU patients	Not measured/NR	Not measured/NR	Delirium presence	IGF-1	Blood	Radioimmunoassay	Age, severe sepsis and APACHE II			IGF-1	
Van der Boogaard et al. (2011) a [69]	100	ICU patients <sup>a</sup>	Not measured/NR	Not measured/NR	Delirium presence	TNF-α, IL-1β, IL-6, IL-8, IL-17, IL-18, MIF, IL-1RA, IL-10, MCP-1, HNP-1, CRP, PCT, Ab1-42, Ab1-40, S100β, cortisol	Plasma	Luminex assay, immunologic detection, and an immunometric assay	NR	<b>Delirium vs non-delirium:</b> IL-8 <sup>a</sup> , IL-10 <sup>g</sup> , Ratio Ab1-42/Ab1-40, TNF-α, IL-6, MIF, IL-1RA, MCP-1, PCT, cortisol, Ab1-40/N-40, Ratio Ab1-42/N-42, Ratio Ab1-40/N-40	<b>Delirium vs non-delirium:</b> IL-17, IL-18, HNP, CRP, S100β, Tau, Ratio Tau/Ab1-40, Ab1-40, Ratio Ab1-40/Ab1-42, Ratio Ab1-40/N-40		<b>Inflamed delirium vs non-inflamed delirium:</b> IL-1β, IL-6, MIF, IL-10, cortisol, ABN-42, IL-18, IL-17, HNP, S100β, Tau, tau/Ab1-40, Ratio Tau/Ab1-40, Ab1-40, Ratio Ab1-40/Ab1-42, Ratio Ab1-40/N-40
Van der Boogaard et al. (2011) b [70]	20	ICU patients	Not measured/NR	Not measured/NR	Delirium presence	Proteomics <sup>h</sup>	Urine + Blood	NR	No multivariate analysis			CRP, Cre	
Burkhardt et al. (2010) [71]	113	Patients aged 265 undergoing elective cardiac surgery with CPB	Not measured/NR	Not measured/NR	Delirium presence	CRP	NR	NR	EuroSCORE, Leucocytes, CRP max, Fentanyl intraoperatively, duration of mechanical ventilation, packed RBC, and treated PONV	CRP		None	
Mu et al. (2010) [72]	243	Patients undergoing elective CABG surgery	Not measured/NR	Not measured/NR	Delirium incidence	Cortisol	Serum	CLIA	Age, history of diabetes mellitus, PRE-OP LVEF, PRE-OP NYHA, preop EuroSCORE score, duration of surgery, POST-OP APACHE II, serum cortisol, POST-OP LVEF, POST-OP complications (within 1 day)	Cortisol		None	



**Table 1** Characteristics of assays and main findings of included delirium studies\* (Continued)

Author and year	Participants		Endpoints	Biomarkers studied	Biological material	Assay method	Covariates accounted for in multivariate analysis	Results	
	Total (N)	Sample						Number of delirium with cancer/total number delirium (%)	Positive association with at least one delirium endpoint **
Pearson et al. (2010) [73]	20	Patients ≥60 with acute hip fracture awaiting surgery <sup>a</sup>	Delirium presence	Cortisol	CSF + serum	ELISA	No multivariate analysis	Cortisol	None
Plaschke et al. (2010) [74]	114 <sup>a</sup>	Patients undergoing elective CABG <sup>a</sup>	Delirium incidence	Cortisol, IL-6	Plasma	ELISA	No multivariate analysis	IL-6, cortisol	None
Tsuta et al. (2010) [75]	103	ICU patients <sup>a</sup>	-Delirium incidence -Delirium prevalence	CRP	Serum	Immunoturbidimetry	Age, APACHE II, coexistence of infection, use of a mechanical ventilator and length of ICU stay	CRP	None
Van Munster et al. (2010) [76]	120	Patients ≥65 admitted for hip fracture surgery	Delirium presence	Cortisol, IL-6, IL-8, S100B	Plasma	CBA	Age, infection, pre-existent cognitive and functional impairment	Cortisol, IL-6, IL-8, S100B	None
Adams et al. (2009) [77]	67	Patients aged ≥70 admitted to elderly care unit	-Delirium incidence -Delirium severity	APOE, IL-1α, IL-1β, IL-1RA, IL-6, TNF-α, IGF-1, IFN-γ, LIF	Serum	ELISA	No multivariate analysis	IGF-1, IFN-γ, IL-1RA	APOE, IL-1α, IL-1β, IL-6, TNF-α, LIF
Van Munster et al. (2009) [78]	120	Patients ≥65 admitted for hip fracture surgery	Delirium incidence	S100B, NSE	Blood	ECLIA	No multivariate analysis	S100B	NSE
Lenstra et al. (2008) [79]	68	Patients undergoing surgery for hip fracture	Delirium incidence	CRP, IL-6, IGF-1	Blood	ELISA	No multivariate analysis	None	CRP, IL-6, IGF-1
Pfister et al. (2008) [80]	16 <sup>b</sup>	Patients with sepsis	Sepsis-related delirium presence	CRP, IL-6, S-100B, cortisol	Serum	Solid-phase enzyme-labelled chemiluminescent sequential immunometric assay	No multivariate analysis	CRP, S100B, Cortisol	IL-6
Rudolph et al. (2008) [81]	42	Patients undergoing cardiac surgery	Delirium incidence	IL-1β, IL-1RA, IL-6, IFN-α, TNF-α, TNF-R1, TNF-R2, IL-2, IL-2R, IL-7, IL-12p40, p70, IL-15, IFN-γ, IP-10, IL-4, IL-5, IL-10, IL-13, MIP-1a, MIP-1b, MIG, Eotaxin, RANTES, CCL-2, IL-8, GM-CSF, IL-17, DRS	Serum	ELISA	No multivariate analysis	MIP-1a, MIP-1b, MIG, Eotaxin, RANTES, CCL-2	IL-1β, IL-1RA, IL-6, IFN-α, TNF-α, TNF-R1, TNF-R2, IL-2, IL-2R, IL-7, IL-12p40, p70, IL-15, IFN-γ, IP-10, IL-4, IL-5, IL-10, IL-13, IL-8, GM-CSF, IL-17, DRS
Van Munster et al. (2008) [82]	98	Patients ≥65 admitted for hip fracture surgery	Delirium presence	IL-6, IL-8, IL-12 (TNF-α, IL-1β, and IL-10 excluded from analysis)	Plasma	CBA	No multivariate analysis	IL-6, IL-8	IL-12
Adams et al. (2007) [83]	164	Acutely ill patients admitted to elderly care unit	-Delirium prevalence -Delirium resolution	APOE, IL-1α, IL-1β, IL-1RA, IL-6, TNF-α, IGF-1, IFN-γ, LIF, CRP	Serum	ELISA	LogAPACHE II, DRS, CRP, Gender, TNF-α, IFN-γ, IGF-1, IL-1RA, and possession of APOE epsilon 4 allele	IGF-1, APOE, IFNγ	IL-6, IL-1α, IL-1β, IL-1RA, TNF-α, LIF, CRP
de Rooij et al. (2007) [84]	185	Patients aged ≥65 admitted to the Department of Medicine	Delirium (14)	IL-1β, IL-6, IL-8, IL-10, TNF-α, CRP	Serum	CBA	Age, cognitive impairment, and infection	IL-6, IL-8	IL-1β, IL-10, TNF-α, CRP
Plaschke et al. (2007) [85]	37	ICU patients	Delirium presence	SAA, IL-6	Blood	ELISA	No multivariate analysis for IL-6	None	SAA, IL-6
White et al. (2005) [86]	283	Patients ≥75 from emergency medical admissions	-Delirium prevalence -Delirium incidence	CRP, Alb, AChE, BuChE, Aspirin esterase, Benzoylcholinesterase	Plasma	ELISA	No multivariate analysis	CRP, Alb, AChE, BuChE, Aspirin esterase, Benzoylcholinesterase	None
Wilson et al. (2005) [87]	100	Patients ≥75 suffering from significant	Delirium incidence	IGF-1	Plasma	CLIA	Depression, IGF-1 levels and IQCODE scores	IGF-1	None

**Table 1** Characteristics of assays and main findings of included delirium studies\* (Continued)

Author and year	Participants	Endpoints	Biomarkers studied	Biological material	Assay method	Covariates accounted for in multivariate analysis	Results
Belooesky et al. (2004) [88]	Total (N) Sample 32 Patients undergoing surgery for hip fracture	Number of delirium with cancer/total number delirium (%) Not measured/NR	CRP, FBG	Blood	Nephelometric assay	Unclear	Positive association with at least one delirium endpoint **
Robertsson et al. (2001) [89]	172 Patients <80 referred to the neuropsychiatric diagnostic unit with suspected dementia	Not measured/NR	Cortisol	Serum	NR	Age, severity of dementia and severity of delirium	Cortisol
Van der Mast et al. (2000) [90]	296 <sup>k</sup> Patients admitted for elective cardiac surgery	Not measured/NR	Try, Ile, Val, Met, Leu, Tyr, Phe, Ser, cortisol	Plasma	HPLC	Plasma amino acids; the ratios of Trp/olNAAs, Tyr/olNAAs, and Phe/olNAAs; albumin; cortisol; and thyroid functions.	Trp, Trp/olNAAs Cortisol, Ile, Val, Met, Leu, Tyr, Phe, Ser
Van der Mast et al. (1999) [91]	296 Patients admitted for elective cardiac surgery	Not measured/NR	Alb, cortisol, 5-HT, try, phe, val, ileu, ile, tryptophane	Plasma	HPLC	Age, inclusion as an inpatient; use of heparine; MMSE score; GFD score; DAL score; Albumin; ratio 113:15; ratio Phe:olNAAs	Alb, phe: ile, Phe:Leu, Phe:val, Phe:tyr, Phe: try
Gustafson et al. (1993) [92]	155 Stroke patients	Not measured/NR	Cortisol	Plasma	Radioimmunoassay	Intercept, basal plasma cortisol, paresis, age, left-sided brain lesion, sex, anti-cholinergic medication, post-decubathesone plasma cortisol	Cortisol
McIntosh et al. (1985) [93]	7 Male patients admitted to hospital for elective surgery	Not measured/NR	Cortisol, B-endorphin	Plasma	Radioimmunoassay	No multivariate analysis	Cortisol, B-endorphin

\* Studies with both delirium and cancer participants are bolded; red coloured biomarkers indicate significance in multivariate analysis

<sup>a</sup> Dementia was an exclusion criteria

<sup>b</sup> Only CRP is reported from this study

<sup>c</sup> Only between incident and prevalent delirium

<sup>d</sup> Pre-operative and post-operative cortisol remained significantly increased in delirium, however, after controlling for pre-operative depression, only preoperative cortisol concentration remained significant, irrespective of the cortisol level after surgery.

<sup>e</sup> Only 66 included in the primary analysis

<sup>f</sup> In inflamed patients only

<sup>g</sup> In non-inflamed patients only

<sup>h</sup> Only CRP and Cre are reported

<sup>i</sup> Same cohort as Plaschke et al. 2007

<sup>j</sup> Only 16 were analysed

<sup>k</sup> same cohort as Van Der Mast et al. 1999

Abbreviations: 5-HIAA 5-Hydroxyindoleacetic acid, 5-HT Serotonin, 6-SMT 6-sulfatoxymelatonin, 8-iso PGF2a 8-iso-prostaglandin F2a, A1A Alpha-1 antitrypsin, a-1-AGP a-1-acid glycoprotein, AA Anticholinergic activity, AB1 Amyloid-B, ACHE Acetylcholinesterase, ACS Acute Coronary Syndrome, ADAS Alzheimer's Disease Assessment Scale, ADL Activities of daily living, Ala Alanine, Alb Albumin, AD Alzheimer's Disease, APACHE Acute Physiology and Chronic Health Evaluation, APN Adiponectin, ANG Angiotensin, APOA1 Apolipoprotein A1, APOE Apolipoprotein E, Arg Arginine, APS Acute Physiology Score, ASA American Society of Anesthesiologists Scale, BCA The bicinchoninic acid assay, BDNF Brain-Derived Neurotrophic Factor, BH4 Tetrahydrobiopterin, BJ B-Endorphin-Like Immunoreactivity, BuCHE Butyrylcholinesterase, C3 Complement C3, CABG Coronary Artery Bypass Graft, CBA Cytometric bead array immunoassay, CCT Charlson Comorbidity Index, CK Creatine Kinase, CK-MB Creatine Kinase-MB, CIA Chemiluminescence immunoassay, CMTN-1 Contactin-1, CPB Cardiopulmonary Bypass, Cre Creatinine, CRP C-Reactive Protein, E2 Estradiol, FBG Fibrinogen, FBN-1 Fibulin-1, ECLIA Electrochemiluminescence immunoassay, EGF Epidermal Growth Factor, FGF-2 Fibroblast Growth Factor, Fk-37 FMS-like tyrosine kinase 3 ligand, GABA Gamma-Aminobutyric Acid, G-CSF Granulocyte Stimulating Factor, GFAP Glial Fibrillary Acidic Protein, GHQ General Health Questionnaire, Glu Glutamic acid, Gly Glycine, GM-CSF Granulocyte-Macrophage Colony-Stimulating Factor, HADS Hospital Anxiety and Depression Scale, Hb Haemoglobin, HCY Homocysteine, HNP-1 Defensin, HP Haptoglobin, HPLC High-performance liquid chromatography, HVA Homovanillic Acid, IADL Instrumental activities of daily living, ICU Intensive care unit, Ile Isoleucine, CAM-1 Interleukin-1 Receptor Antagonist, Ile Isoleucine, IP-10 Interferon gamma-induced protein 10, IJCODOE The Informant Questionnaire on Cognitive Decline in the Elderly, KYN Kynurenine, Leu Leucine, LIF Leukaemia Inhibitory Factor, LMAA Large Neutral Amino Acids, LOS Length of stay, LP Leptin, Met Methionine, MB-CX MB-isoform of Creatinine Kinase, MCP Monocyte Chemoattractant Protein, MPO Myeloperoxidase, MT Melatonin, MCM Neural Cell Adhesion Molecule, MIF Macrophage Migration Inhibitory Factor, MIP Macrophage Inflammatory Protein, MMP-9 Matrix Metalloproteinase-9, MMSE Mini-mental state examination, MPO Myeloperoxidase, MVAH New York Heart Association, PACU Post-anesthesia care unit, PAI-1 Plasminogen activator inhibitor-1, ACT Procalcitonin, PDGF Platelet-Derived Growth Factor, Phe Phenylalanine, pW/PC Plasma NF Not reported, NSE Neuron Specific Enolase, Orn Ornithine, NYHA New York Heart Association, PO22 Post-operative, PO202 Post-operative, PO2022 Post-operative, day 2, PONY Post-operative nausea and vomiting, POST-OP Post-operative, PRE-OP Pre-operative, P-tau Phosphorylated tau, RANTES Chemokine (C-C motif) ligand 5, RBC Red blood cell, S100B s100 calcium-binding protein B, sCD40L Soluble CD40 ligand, Ser Serine, sIL-XR Soluble IL-X receptor, SJJ Somatostatin-Like Immunoreactivity, STNER Soluble Tumor Necrosis Factor Receptor, Tau Taurine, T-tau Total tau, TGF-α Transforming Growth Factor Alpha, THA Total Hip Arthroplasty, TRACE Time Resolved Amplified Cryptate Emission, TSH Thyroid Stimulating Hormone, TNF Tumor Necrosis Factor, Trp Tryptophan, TRX Thioresoxin, Tyr Tyrosine, UDL Under detection limit, Val Valine, VCAM-1 Vascular Cell Adhesion protein 1, VEGF Vascular Endothelial Growth Factor, wWF Von Willebrand factor, ZAG Zinc-z-2-Glycoprotein

**Table 2** Characteristics of assays and main findings of included cancer studies\*

Author and year	Participants		Endpoints	Biomarkers studied	Biological material	Assay method	Covariates adjusted for in multivariate analysis	Results	
	Total participants (N)	Cases; control						Positive association with at least one endpoint**	Negative association
Amano et al. (2017) <sup>a</sup> [94]	1702	Advanced cancer patients; no control	-Anorexia -Weight loss -Fatigue -Dyspnea -Dysphasia -Edema -Pressure ulcer -ADL disabilities	CRP	NR	NR	Age, gender, primary tumor site, distant metastasis, chemotherapy, ECOG PS, and setting of care	CRP	None
Demiray et al. (2017) [95]	87	Participants with advanced cancer; healthy participants without a known chronic disease	-Cachexia -Weight loss -PFS -OS	LP, resistin	Serum	ELISA	NR	Multivariate results NR	LP Resistin* Multivariate results NR
Fogelman et al. (2017) [96]	69	Participants with advanced cancer; healthy controls with no cancer diagnosis	Either 10% weight loss or death at 60 days from the start of therapy	APN, bFGF, CXCL-16, FSN, Ghrelin, IGF-1, IL-1 $\beta$ , IL-6, IL-8, Klotho, LP, MCP-4, MK, MSTN, PIF, sTNFR1, sTNFR2, TARC, TNF- $\alpha$ , VEGF, ZAG	NR	NR	Smoking status, best response, pain, difficulty swallowing	MK, IL-1 $\beta$ , CXCL-16, IL-6, IL-8, TNF- $\alpha$ Multivariate results NR	APN, bFGF, FSN, Ghrelin, IGF-1, Klotho, LP, MCP-4, MSTN, MK, PIF, sTNFR1, sTNFR2, TARC, VEGF, ZAG Multivariate results NR
Luo et al. (2017) [97]	217	Participants with advanced cancer; no control	-PFS -OS	FBG, CA-125, NLR, PLR	Serum + Plasma	NR	NR	FBG	CA-125, NLR, PLR
Paulsen et al. (2017) [98]	49	Participants with cancer; no control	-Pain -Appetite -Fatigue	CRP, ESR, sTNF-R1, IL-1RA, IL-6, MCP-1, IL-18, MIF, TGF- $\beta$ 1	Serum	ELISA (multiplex assay)	Sex, BMI and age	sTNF-r1, MCP-1, MIF, CRP, IL-6, IL-1RA	IL-18, TGF- $\beta$ 1, ESR
Amano et al. (2016) [99]	1511	Advanced cancer patients; no control	-Survival rate -Mortality rate	CRP	Plasma	Latex-enhanced immunoturbidimetric assay	Age, gender, primary tumor site, distant metastasis, chemotherapy, ECOG PS, and setting of care	CRP	None
Bye et al. (2016)	60	Participants	-Cachexia	IL-10, IFN- $\gamma$ , LP, APN,	Serum	ELISA	No multivariate	IL-6	IL-10, IFN- $\gamma$ ,

**Table 2** Characteristics of assays and main findings of included cancer studies\* (Continued)

Author and year	Participants		Endpoints	Biomarkers studied	Biological material	Assay method	Covariates adjusted for in multivariate analysis	Results	Negative association
	Total participants (N)	Cases; control							
[100]		with advanced cancer; healthy controls with normal weight	-Survival	TNF- $\alpha$ , IL-6, IGF-1			analysis		TNF- $\alpha$ , APN, IGF-1
Mitsunga et al. (2016) [101]	421	Participants with advanced cancer with low, intermediate and high CRP levels	OS	CRP, NLR	Blood	ELISA (Multiplex assay)	<b>Retrospective cohort:</b> Sex, age, ECOG-PS, UICC stage, CA 19-9, prognostic CRP classification; <b>Prospective cohort:</b> Sex, age, ECOG-PS, UICC stage, CA 19-9, NLR classification, mGPS, prognostic CRP classification	CRP, NLR	None
Morgado et al. (2016) [102]	49	Participants with advanced cancer and fatigue with and without weight loss	-Weight loss -Fatigue	Hb, LDH, Alb, CRP, Cre	Serum + Urine	NR	No multivariate analysis	Alb, CRP	Hb, LDH, Cre
Rodrigues et al. (2016) [103]	51	Participants with advanced cancer; no control	Fatigue	IL-1, IL-6, TNF- $\alpha$ , $\alpha$ -1-AGP, GPS (Alb+CRP)	Blood	NR	No multivariate analysis	TNF- $\alpha$ , GPS (Alb+CRP)	None
Srdic et al. (2016) [104]	100	Participants with advanced cancer with and without cachexia	-Cachexia -Chemotherapy toxicity -Survival	CRP, IL-6, Alb, Hb	NR	The Bromocresol Purple method	NR	CRP, IL-6, Alb, Hb	None
Wu et al. (2016) [105]	55	Participants with advanced cancer; no control	-OS -PFS	NLR, PLR, ALP, LDH	Blood	NR	NR	PLR, NLR, LDH	ALP
Bilir et al. (2015)	80	Participants	-OS	IL-1 $\beta$ , IL-1 $\alpha$ , IL-6, TNF- $\alpha$	Serum	ELISA	NR	CRP, TRAF-6, Alb, LDH, IL-1 $\alpha$ , IL-6	IL-1 $\beta$

**Table 2** Characteristics of assays and main findings of included cancer studies\* (Continued)

Author and year	Participants		Endpoints	Biomarkers studied	Biological material	Assay method	Covariates adjusted for in multivariate analysis	Results	Negative association
	Total participants (N)	Cases; control							
[106]		with advanced cancer and cachexia; healthy controls with no known chronic disease or weight loss	-Cachexia	orexin-A, galanin, TWEAK, TRAF-6, NPY, CRP, Testosterone, Alb, LDH				TNF- $\alpha$ , TWEAK, orexin-A, NPY, testosterone	galanin
Miura et al. (2015) [107]	79	Participants with advanced cancer; no control	-Body composition -Fatigue	IL-6	Serum	ELISA (multiplex assay)	NR	IL-6	None
Miura et al. (2015) b [108]	1160	Participants with advanced cancer; no control	Survival	mGPS (Alb+CRP)	NR	NR	Primary tumor site, age and gender	mGPS (Alb+CRP)	None
Barrera et al. (2014) [109]	135	Participants with advanced cancer; healthy controls	-Quality of life (fatigue, PS, hyporexia, BMI) -Survival	IL-31, IL-33, IL-27, IL-29, IL-1 $\beta$ , IL-2, IL-6, IL-8, IL-12p70, IL-17A, IFN- $\gamma$ , TNF- $\alpha$ , IL-4, IL-10	Plasma	CBA	No multivariate analysis	IL-6, IL-8, IFN- $\gamma$ , IL-33, IL-10, IL-29 <sup>b</sup> , IL-12p70 <sup>b</sup> , IL17a <sup>b</sup>	IL-31, IL-27, IL-1 $\beta$ , IL-2, TNF- $\alpha$ , IL-4
Blakely et al. (2014) [110]	50	Participants with advanced cancer with normal CRP and elevated CRP	-OS -Mortality rate -gastrointestinal obstruction -Pain -Bleeding -Other symptoms (NR) -Major complications	CRP	Serum	NR	NR	CRP	None
Fujiwara et al. (2014) [111]	21	Participants with advanced cancer with and without cachexia	Cachexia	LP, IL-6, TNF- $\alpha$	Serum	ELISA	No multivariate analysis		LP, IL-6, TNF- $\alpha$
Lindemann et al. (2014)	218	Participants with	-Survival -Weight loss	CRP, Alb	Plasma	Immune-turbidimetry	No multivariate analysis	CRP, Alb	None

**Table 2** Characteristics of assays and main findings of included cancer studies\* (Continued)

Author and year	Participants	Endpoints		Biological material	Assay method	Covariates adjusted for in multivariate analysis	Results	
		Cases	control				Positive association with at least one endpoint**	Negative association
[112]		advanced cancer; no control						
Mondello et al. (2014) [113]	170	Participants with advanced cancer; healthy controls	-Survival -Cachexia	Serum	ELISA	Age, ghrelin, obestatin, leptin, disease and chronic kidney disease	LP, Ghrelin, obestatin	None
Moriwaki et al. (2014) [114]	62	Patients with advanced cancer with GPS 0, GPS 1 or GPS 2	OS	NR	NR	GPS, median ALP, median LDH, number of metastatic organs, liver metastasis, peritoneal metastasis, other metastasis	GPS (Alb+CRP)	ALP, Bilirubin, LDH, CEA, CA 19-9
Szkandera et al. (2014) [115]	474	Participants with cancer; no control	Cancer-specific survival	Plasma	NR	Age, gender, tumour grade, tumour stage, administration of chemotherapy, surgical resection, NLR, PLR, bilirubin levels and plasma CRP levels	CRP, NLR	PLR
Zhang et al. (2014) [116]	200	Participants with cancer; no control	-Fatigue -Chemotherapy adverse effects	Plasma + urine	ELISA	No multivariate analysis	TNF- $\alpha$ , IL-1 $\alpha$ , IL-1 $\beta$ , 17-HCS	17-HCS
Jafri et al. (2013) [117]	173	Participants with advanced cancer with high inflammation and with low inflammation	-PFS -OS	Serum	NR	Sex, race, PS and histology	ALI (Alb+NLR)	None
Laird et al. (2013) [118]	1466	Participants with advanced cancer with low and high CRP levels	-Symptoms of the EOTC (pain, appetite loss, cognitive function, dyspnea, fatigue, physical function,	Blood	NR	No multivariate analysis	CRP	None

**Table 2** Characteristics of assays and main findings of included cancer studies\* (Continued)

Author and year	Participants		Endpoints	Biomarkers studied	Biological material	Assay method	Covariates adjusted for in multivariate analysis	Results	
	Total participants (N)	Cases; control						Positive association with at least one endpoint**	Negative association
Laird et al. (2013) b [119]	2456	Participants with advanced cancer; no control	role function, social function, QoL, nausea/vomiting, diarrhea, sleep, constipation) -Survival -Symptoms of the EOTC (pain, appetite loss, cognitive function, dyspnea, fatigue, physical function, role function, social function, QoL, nausea/vomiting, diarrhea, sleep, constipation) -Survival	mGPS (Alb+CRP)	Blood	NR	NR	mGPS (Alb+CRP)	None
Paiva et al. (2013) [120]	223	Participants with cancer with and without fatigue	-Fatigue -OS	CRP, Hb, LDH, Alb	Blood	NR	Age, KPS, type of treatment, breast cancer, upper gastrointestinal cancer, head and neck cancer, lower gastrointestinal cancer, lung cancer, urologic cancer, and CRP	CRP, Hb, LDH, Alb, WBC	None
Suh et al. (2013) [121]	98	Participants with advanced cancer; no control	Survival	IL-6, TNF- $\alpha$	Plasma	ELISA (multiplex assay)	Gender (male), fatigue (BFI-K score), ECOG (3-4), IL-6 (high, $\geq 9.06$ pg/mL)	IL-6	TNF- $\alpha$
De Raaf et al. (2012) [122]	92	Participants with advanced cancer survivors	Physical and mental fatigue	CRP, IL-1-RA, NP, IL-6 and IL-8	Plasma	CBA	No multivariate analysis	CRP, IL-6, IL-1-ra, NP	IL-8
Gjoulbasanis et al. (2012)	114	Participants with	-Nutritional status (cachexia)	IL-8	Plasma	CLIA	PS, histology, BMI, gender, age,	IL-8	None

**Table 2** Characteristics of assays and main findings of included cancer studies\* (Continued)

Author and year	Participants		Endpoints	Biomarkers studied	Biological material	Assay method	Covariates adjusted for in multivariate analysis	Results	Negative association
	Total participants (N)	Cases; control							
[123]		advanced cancer with malnutrition, with a risk of malnutrition, and who were well nourished	-Survival				smoking status, weight loss history		
Gulen et al. (2012) [124]	88	Participants with advanced cancer with and without weight loss; age- and sex-matched controls	Weight loss (>5%)	LP, APN, TNF- $\alpha$ , CRP	Serum	ELISA	No multivariate analysis	LP	APN, TNF- $\alpha$ , CRP
Heitzer et al. (2012) [125]	65	Advanced cancer patients with cancer pain; healthy controls without pain	Pain intensity	IL-1 $\beta$ , IL-2, IL-4, IL-5, IL-6, IL-8, IL-10, IL-12, TNF- $\alpha$ , TNF- $\beta$ , IFN- $\gamma$ , IL-1 $\alpha$ , IL-7, IL-13, IL-18, MCP-1, MIP-1 $\alpha$ , MIP-1 $\beta$ , OPG	Serum	ELISA	NI	Unclear	Unclear
Minton et al. (2012) [126]	720	Participants with advanced cancer with and without fatigue	Fatigue	CRP, Alb, Hb	Blood	NR	Hb, current treatment with chemo, QOL score, depression, pain dyspnoea, cognitive function, insomnia and loss of appetite	CRP, Alb, Hb	None
Partridge et al. (2012) [127]	102	Patients with advanced cancer with GPS 0, GPS 1 or GPS 2 ; no control	Survival	mGPS (Alb+CRP)	Blood	NR	Sex, primary cancer site, age, Hb and WBC	mGPS (Alb+CRP)	None
Pond et al. (2012) [128]	220	Participants with advanced cancer; no control	-OS -PFS	CRP	NR	NR	NR	CRP	None
Wang et al.	177	Participants	Survival	CRP, Alb, mGPS	NR	NR	PS, pretherapeutic	CRP, mGPS (Alb+CRP), NLR	Alb



**Table 2** Characteristics of assays and main findings of included cancer studies\* (Continued)

Author and year	Participants	Endpoints		Biological material	Assay method	Covariates adjusted for in multivariate analysis	Results	
		Cases	control				Positive association with at least one endpoint**	Negative association
(2012) [129]		with cancer;		(Alb+CRP), NLR		weight, WBC, neutrophil count, NLR, CRP, mGPS, PI, the 7 <sup>th</sup> TNM staging, surgery, degree of differentiation, palliate chemotherapy		
Aydin et al. (2011) [130]	61	Advanced cancer patients; no control	Survival	Serum	Nephelometric assay	No multivariate analysis	CRP, Alb, TFN	None
Dev et al. (2011) [131]	77	Participants with advanced cancer; no control	Symptom distress (pain, fatigue, nausea, depression, anxiety, drowsiness, appetite, well-being, dyspnea, sleep)	Serum	NR	NR	Cortisol	None
Gioulbasanis et al. (2011) [132]	115	Participants with advanced cancer with malnutrition, with a risk of malnutrition, and who were well nourished	-Nutritional status (cachexia) -Survival	Plasma	Radioimmunoassay	Number of metastatic sites, PS, weight loss <5%, MNA groups, age, and major histological type	CRP, LP, Alb	Ghrelin, APN, IGF-1
Hwang et al. (2011) [133]	402	Participants with cancer; no control	-PFS -OS	Serum	Latex turbidimetric immunoassay	Peritoneal metastasis, bone metastasis, albumin, CRP, ECOG PS, GPS	Alb, CRP	None
Kwak et al. (2011) [134]	90	Participants with advanced cancer; no control	Fatigue	Blood	NR	BFI score, age, gender, BMI, blood pressure, heart rate, cancer site, previous treatment, comorbidity,	IL-6, TNF- $\alpha$	None

**Table 2** Characteristics of assays and main findings of included cancer studies\* (Continued)

Author and year	Total participants (N)	Participants		Endpoints	Biomarkers studied	Biological material	Assay method	Covariates adjusted for in multivariate analysis	Results	Negative association
		Cases	control							
Lee et al. (2011) [135]	126	Participants with advanced cancer; no control	Participants with advanced cancer; no control	14 day mortality	CRP	Serum	NR	CRP, chemotherapy, age, dyspnea, altered mental status, hypotension, and leukocytosis	CRP	None
Scheede-Bergdahl et al. (2011) [136]	83	Participants with advanced cancer; no control	Participants with advanced cancer; no control	- Clinical features of cachexia (weakness, loss of appetite, fatigue, QOL, weight loss) -Survival	IL- 6, IL-1 $\beta$ , IL-8, TNF- $\alpha$	Plasma	BCA	Sex, age, diagnosis, oncological treatment, CCI and medications	IL- 6, IL-1 $\beta$ , IL-8, TNF- $\alpha$	None
Vlachostergios et al. (2011) [137]	77	Participants with advanced cancer; no control	Participants with advanced cancer; no control	-TTP -OS	IGF-1, CRP, Alb	Serum	Radioimmunoassay	Sex, current smoker, albumin, IGF-1	IGF-1, CRP, Alb	None
Diakowska et al. (2010) [138]	218	Participants with cancer with and without cachexia, healthy blood donors; and patients with non-malignant diseases of alimentary tract	Participants with advanced cancer; no control	Cachexia	LP, CRP, IL-1, IL-6, IL-8, TNF- $\alpha$ , Alb, Hb.	Serum	ELISA	NR	LP, IL-6, Alb, TNF- $\alpha$	IL-1, IL-8, Hb, CRP*
Meek et al. (2010) [139]	56	Participants with advanced cancer; no control	Participants with advanced cancer; no control	Cancer-specific survival	IGF-1, IGFBP-3, CRP, mGPS (Alb+CRP), LP	Serum	NR	BMI, cancer stage, Hb, WBC, mGPS	mGPS (Alb+CRP)	IGF-1, IGFBP-3, LP, CRP

**Table 2** Characteristics of assays and main findings of included cancer studies\* (Continued)

Author and year	Total participants (N)	Endpoints		Biomarkers studied	Biological material	Assay method	Covariates adjusted for in multivariate analysis	Results	
		Cases	control					Positive association with at least one endpoint**	Negative association
Ishizuka et al. (2009) [140]	112	Participants with advanced cancer; no control	Mortality	CRP, Alb, mGPS (Alb+CRP), Neutrophil ratio	Serum	NR	Neutrophil ratio, CA 19-9, CRP, albumin, and mGPS	mGPS (Alb+CRP)	None
Karapanagiotou et al. (2009) [141]	161	Participants with advanced cancer; healthy controls	-Weight loss -TTP -OS	Ghrelin, LP	Serum	ELISA	Sex, age, BMI, Ghrelin	Ghrelin Multivariate results NR	LP Multivariate results NR
Paddison et al. (2009) [142]	44	Participants with advanced cancer; healthy controls	Fatigue	Hb, WBC, Neutrophil, Monocyte, Lymphocyte	Blood	NR	Age, gender, time until treatment termination; and fatigue	Hb, WBC, Neutrophil count, monocyte count	None
Takahashi et al. (2009) [143]	26	Participants with cancer cachexia; healthy controls	Anorexia (cachexia and BMI)	TNF- $\alpha$ , IFN- $\gamma$ , IL-6, IL-1RA, LP, ghrelin	Plasma	ELISA	No multivariate analysis	TNF- $\alpha$ , IL-6, IL-1RA, LP	IFN- $\gamma$ , ghrelin
Inagaki et al. (2008) [144]	46	Participants with advanced cancer with and without fatigue	Fatigue	IL-6	Plasma	ELISA	Logistic regression: IL-6, gender, weight and clinical fatigue Multiple regression: gender, weight, IL-6 and total score of the CFS	IL-6	None
Karapanagiotou et al. (2008) [145]	152	Participants with advanced cancer; healthy controls	-Weight loss -TTP -OS	LP, APN, resistin	Serum	ELISA	Sex, age, BMI, resistin	Resistin	LP, APN
Sharma et al. (2008) [146]	52	Participants with advanced cancer; no control	-OS -Toxicity	IL-1 $\beta$ , IL-2, IL-4, IL-5, IL-8, IL-6, IL-10, IL-12, GM-CSF, IFN- $\gamma$ , TNF- $\alpha$ , sIL-6R, sgp130, VEGF, eotaxin, MCP-1, MIP-	Serum	NR	Tumour site (colonic primary), GPS, CEA, and albumin	GPS (Alb+CRP), Hb, Alb	CRP, IL-1 $\beta$ , IL-2, IL-4, IL-5, IL-8, IL-6, IL-10, IL-12, GM-CSF, IFN-

**Table 2** Characteristics of assays and main findings of included cancer studies\* (Continued)

Author and year	Participants	Endpoints		Biological material	Assay method	Covariates adjusted for in multivariate analysis	Results	Negative association
		Cases	control					
Weyńska et al. (2008) [147]	40	Participants with advanced cancer with and without cachexia	- Cachexia - Nutritional status	Serum	ELISA	No multivariate analysis	LP	Y, TNF-α, sIL-6R, sgp130, VEGF, eotaxin, MCP-1, MIP-1α, MIP-1β None
Ravasco et al. (2007) [148]	101	Participants with cancer; no control	-REE -Weight loss -Nutritional intake	Serum	ELISA	Cancer histology and stage, nutritional intake	IL-1RA, IL-6, TNF-α, IFN-γ, VEGF	IL-10
Richey et al. (2007) [149]	24	Participants with cancer with and without cachexia	Cachexia	Serum	Dry-slide method with the VITROS Fusion Series analyser	No multivariate analysis	GPS (Alb+CRP), Alb, CEA	IL-1α, IL-1β, IL-2, IL-4, IL-5, IL-6, IL-8, IL-10, IL-12, TNF-α, IFN-γ, VEGF, GM-CSF, MCP-1, MIP-1a, MIP-1B, RANTES, FGF, Hb, CRP, CEA
Suh et al. (2007) [150]	44	Participants with advanced cancer; no control	Survival	Serum	NR	NR	CRP	None
Al Murri et al. (2006) [151]	96	Breast cancer patients; no control	Survival	NR	NR	GPS and treatment	CRP, GPS (Alb + CRP)	None
Kayacan et al. (2006) [152]	56	Participants with advanced cancer with and without cachexia; healthy smokers for the control	- Cachexia -PS -Survival	Serum	ELISA	NR	None	TNF-α, IL-6

**Table 2** Characteristics of assays and main findings of included cancer studies\* (Continued)

Author and year	Participants		Endpoints	Biomarkers studied	Biological material	Assay method	Covariates adjusted for in multivariate analysis	Results	
	Total participants (N)	Cases; control						Positive association with at least one endpoint**	Negative association
Ramsey et al. (2006) [153]	119	Participants with advanced cancer; no control	-Cancer-specific survival -Cancer-specific mortality	GPS (Alb+CRP)	NR	NR	GPS, Hb, calcium, WBC, neutrophil count, Alb, CRP	GPS (Alb+CRP)	None
Di Nisio et al. (2005) [154]	141	Participants with advanced cancer; no control	Survival	IL-6, IL-10, IFN- $\gamma$ , P-selectin	Plasma	BCA	Life expectancy, WHO performance status, concomitant treatment, type of carcinoma, and histology	IL-10, IL-6, P-selectin	IFN- $\gamma$
Rich et al. (2005) [155]	80	Participants with advanced cancer with good and dampened circadian rhythms	-Extent of metastatic disease -PS -QOL	IL-6, TGF- $\alpha$ , TNF- $\alpha$ , cortisol	Serum	ELISA	NR	IL-6, TGF- $\alpha$ , TNF- $\alpha$	Cortisol
Bolukbas et al. (2004) [156]	69	Participants with advanced cancer; healthy controls with stable weight	Weight loss	LP	Serum	ELISA	NR	LP	None
De Vita et al. (2004) [157]	68	Participants with advanced cancer; no control	-TTP -OS	IL-6	Serum	ELISA	NR	IL-6	None
Dulger et al. (2004) [158]	54	Participants with advanced cancer with and without cachexia; healthy gender- and age-matched adults	Cachexia	TNF- $\alpha$ , IL-1 $\beta$ , IL-6, CRP, LP, GH, TG, insulin, glucose, triglyceride, total protein, ESR	Serum	Solid-phase, two-site chemiluminescent immunoassays	No multivariate analysis	Alb, total protein, GH, TNF- $\alpha$ , IL-1 $\beta$ , IL-6, insulin, LP, ESR <sup>p</sup> , CRP <sup>b</sup>	Glucose, TG
Elahi et al.	165	Participants	Survival	Alb, CRP	NR	Fluorescence	NR	Alb, CRP	None

**Table 2** Characteristics of assays and main findings of included cancer studies\* (Continued)

Author and year	Participants		Endpoints	Biomarkers studied	Biological material	Assay method	Covariates adjusted for in multivariate analysis	Results	Negative association
	Total participants (N)	Cases; control							
(2004) [159]		with advanced cancer; no control				polarization immunoassay			
Jamieson et al. (2004) [160]	33	Participants with advanced cancer; healthy controls	Weight loss	Hb, Alb, CRP, APN, LP, IL-6	Serum	ELISA	No multivariate analysis	Hb, Alb, CRP, APN, LP, IL-6	None
Songur et al. (2004) [161]	91	Participants with advanced cancer; healthy controls	-Malnutrition -Survival	IL-6, Alb, CRP, TFN, LDH	Serum	NR	NR	IL-6, Alb, CRP, TFN, LDH	None
Scott et al. (2003) [162]	106	Participants with advanced cancer with and without weight loss	-Weight loss	Hb, Alb, CRP	Blood	NR	No multivariate analysis	Hb, Alb, CRP	None
Aleman et al. (2002) [163]	106	Patients newly diagnosed with NSCL vs patients with no cancer	-Nutritional status -Survival	IL-6, IL-12, IL-10, IL-2, LP, $\alpha$ -1A, ferritin, CRP, TNF- $\alpha$ , s-TNFR2, s-IL-2R, IFN- $\gamma$	Serum	CLIA	NR	IL-6, IL-12, IL-2, sTNFR2, IFN- $\gamma$ , sIL-2R, LP, $\alpha$ -1A, CRP, ferritin Multivariate results unclear	IL-10, TNF- $\alpha$ Multivariate results unclear
Orditura et al. (2002) [164]	85	Participants with advanced cancer; healthy controls	-OS -TTF	IL-8, IL-10, IL-2	Serum	ELISA	NR	IL-10, IL-2, IL-8	None
Scott et al. (2002) [165]	106	Participants with advanced cancer; no control	Survival	Alb, CRP	Blood	NR	Age, sex, stage, histological type, weight loss, haemoglobin, albumin, CRP, KPS and EORTC QLQ-C30 subscale	CRP, Alb	None
Jatoi et al. (2001) [166]	73	Participants with	Anorexia and/or weight loss	NPY, LP, CCK-8	Serum	Radioimmunoassay	No multivariate analysis	NPY	LP, CCK-8

**Table 2** Characteristics of assays and main findings of included cancer studies\* (Continued)

Author and year	Participants		Endpoints	Biomarkers studied	Biological material	Assay method	Covariates adjusted for in multivariate analysis	Results	
	Total participants (N)	Cases; control						Positive association with at least one endpoint**	Negative association
Mantovani et al. (2001) [167]	58	Participants with advanced cancer; healthy controls	-BMI -Cachexia -ECOG PS -Survival	LP, IL-6, TNF- $\alpha$	Serum	ELISA	No multivariate analysis	Unclear	Unclear
Mantovani et al. (2000) [168]	32	Participants with advanced cancer; normal weight healthy controls	-cachectic symptoms (BMI)	LP, IL-1 $\alpha$ , IL-6, and TNF- $\alpha$	Serum	ELISA	No multivariate analysis	Unclear	Unclear
Nenova et al. (2000) [169]	87	Participants with advanced cancer; healthy controls	-Cachexia -Prognosis	TNF- $\alpha$	Serum	ELISA	No multivariate analysis	Unclear	Unclear
O'Gorman et al. (1999) [170]	50	Participants with advanced cancer with weight loss or weight gain; weight stable controls	-Weight loss -Appetite -PS -Inflammation	Alb, CRP	Blood	NR	No multivariate analysis	Alb, CRP	None
Okada et al. (1998) [171]	100	Participants with cancer; healthy controls	Weight loss	IL-6	Serum	ELISA	No multivariate analysis	IL-6	None
Wallace et al. (1998) [172]	54	Participants with advanced cancer; healthy	Weight loss	LP	Serum	Radioimmunoassay	No multivariate analysis	LP	None

**Table 2** Characteristics of assays and main findings of included cancer studies\* (Continued)

Author and year	Participants		Endpoints	Biomarkers studied	Biological material	Assay method	Covariates adjusted for in multivariate analysis	Results	
	Total participants (N)	Cases; control						Positive association with at least one endpoint**	Negative association
Maltoni et al. (1997) [173]	530	controls Participants with advanced cancer; no control	Survival	Neutrophil, lymphocyte & monocyte %, basophil + eosinophil %, Hb, TFN, Alb, total WBC, Pseudocholinesterase, proteinuria, TFN, transport iron	Blood	NR	No multivariate analysis	Neutrophil %, lymphocyte %, total WBC, CHE, Alb	basophil + eosinophil %, Hb, TFN
Simons et al. (1997) [174]	21	Participants with cancer and weight loss; no control	-Weight loss -Body composition -Appetite -REE	LP	Plasma	ELISA	No multivariate analysis	LP	None

Note: Cancer prognosis was not separated from the other syndromes in the table

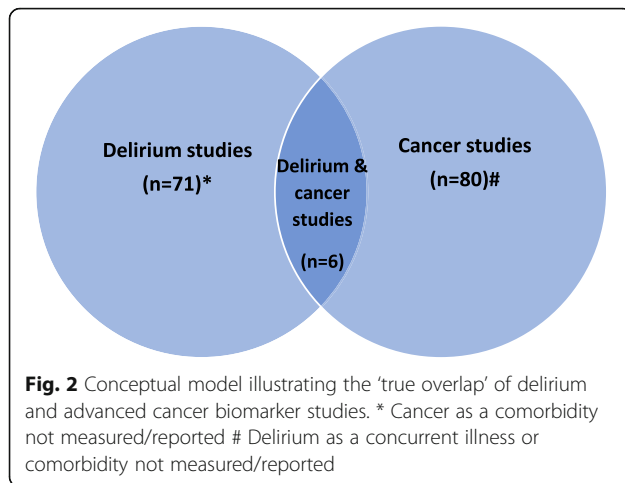
\* Red coloured biomarkers indicate significance in multivariate analysis

<sup>a</sup>Secondary analysis of Amano, 2016

<sup>b</sup>In cancer vs no cancer only

Abbreviations: 17-HCS 17-hydroxycorticosteroids,  $\alpha$ -1-AGP  $\alpha$ -1-acid glycoprotein,  $\alpha$ -1A  $\alpha$ -1 antitrypsin, Alb Albumin, ALP Alkaline phosphatase, APN Adiponectin, APOA2 Apolipoprotein A2, BCA The bichinchoninic acid assay, bFGF Basic fibroblast growth factor, CA 19-9 Cancer antigen, CBA Cytometric bead array immunoassay, CCK Cholecystokinin, CEA Carcinoembryonic antigen, CK Creatine Kinase, CLIA Chemiluminescence immunoassay, Cre Creatinine, CRP C-Reactive Protein, CXCL Soluble CXC chemokine ligand, ESR Erythrocyte sedimentation rate, FBG Fibrinogen, FSN Follistatin, GH Growth Hormone, GM-CSF Granulocyte-Macrophage Colony-Stimulating Factor, HA Hyaluronic Acid, Hb Haemoglobin, IGF Insulin-Like Growth Factor, IGFBP Insulin-like Growth Factor, IGFBP Insulin-like Growth Factor, IL Interleukin, IFN Interferon, LDH Lactate Dehydrogenase, LP Leptin, MCP Monocyte Chemoattractant Protein, MIP Macrophage Inflammatory Protein, MK Midkine, NI Not enough information, NR Not reported, MSTN Myostatin, NLR Neutrophil-lymphocyte ratio, NP Neopterin, NPY Neuropeptide Y, OPG Osteoprotegerin, PLR Platelet-lymphocyte ratio, RANTES Chemokine (C-C motif) ligand 5, sTMFR Soluble Tumor Necrosis Factor Receptor, Sgp130 Soluble glycoprotein 130, TARC Thymus and Activation-Regulated Chemokine, TFN Transferin, TG Triglyceride, TNF Tumor Necrosis Factor, TRAF-6 Tumor Necrosis Factor Receptor associated factor-6, TTF Time to treatment failure, TWEAK TNF-like weak inducer of apoptosis, VEGF Vascular Endothelial Growth Factor, ZAG Zn-alpha2 glycoprotein





participants who had delirium also had cancer, in another two, 26% and 27% of the delirium cohorts had cancer, and in the remaining study 14% of the delirium participants had cancer (Table 1). Although only six delirium studies reported co-existing cancer, there is still uncertainty as to how many participants in both groups of studies had both delirium and cancer. The two most common biomarkers in these six studies that reported a positive association with delirium were CRP ( $n=3$ ) and IL-6 ( $n=3$ ). It is unclear however whether these biomarkers were predominantly associated with delirium or the cancer, as three of the six studies grouped the delirium participants together, irrespective of their cancer comorbidity.

The quality assessment showed a large variability in the reporting of included studies. 150 (99%) studies had a clear aim statement which included their outcome of interest. One study did not report a clear aims statement [175]. One hundred and nineteen studies (79%) did not explicitly state the hypothesis; however, in most ( $n=94$ ; 62%) the hypothesis could be interpreted by the study aim. All 151 studies stated the participant population in detail. No study reported all elements of the assay methods in the REMARK checklist [23]. One hundred and thirty one studies (87%) did not report whether assays were blinded to the study endpoint, however 59 (45%) of those studies were objective assessments. Further, 14 studies (9%) reported a power calculation to justify their sample size. Most ( $n=125$ ; 83%) of studies defined all clinical endpoints examined. Ninety seven (64%) studies undertook multivariate analysis, and of these 67 (69%) described the multivariate model and the covariates included in the model, and 23 (23%) explained the rationale for inclusion of the covariates in the models. (Additional files 4 and 5). Furthermore, 27 delirium studies (38%) did not report the reason for admission. Of the 44 studies that did report the reason for admission, these were predominantly for surgery-

elective and acute ( $n=40$ ). Most studies in the non-surgical population did not report a reason for admission, with the exception of 4 studies where the medical condition of interest occurred on admission (e.g stroke). See additional files 4 and 5 for the complete quality assessments.

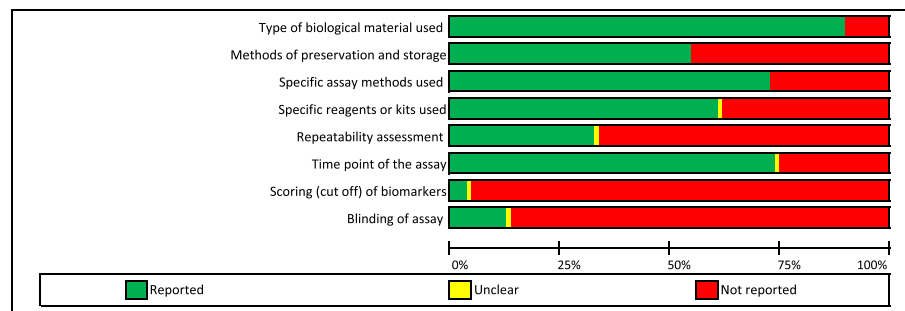
The methodological quality of the assay procedures only is depicted in Figure 3, with reporting of type of biological material mostly provided but much lower frequency of reporting for other critical descriptors.

## Discussion

This is the first systematic review to our knowledge, to demonstrate the high degree of overlap in biomarkers in delirium, cancer prognosis and advanced cancer syndromes. This systematic review of 151 studies found that 41 biomarkers were independently investigated in studies of both delirium and prognosis/advanced cancer syndromes; with over half having a positive association in at least one study.

Biomarkers fall into three categories (though not mutually exclusive); those which present before disease onset that can help identify individuals who are most at risk of a particular disease (for example, genetic markers), those which are disease markers and as such, increase during disease progression and decrease after resolution, and thirdly, biomarker as an end-product of a disease for which levels are proportionate to ‘damage’ due to the disease [176]. The findings of this systematic review suggest that categorization along these lines is less understood in delirium. For example, there is evidence to show that conditions such as sepsis and hip fracture cause changes in inflammatory markers [177, 178], however, there is little evidence about whether delirium self-propagates. Some animal model data in delirium suggests that there might be a direct impact of inflammatory markers on brain dysfunction [179]. To our knowledge there was no published relationship between tumor markers and neurological brain dysfunction. Although clinical evidence suggests long term impacts on brain function, the exact pathophysiological mechanisms are poorly understood, and biomarkers to measure this are also unclear.

The issue of biomarker overlap between associated conditions has been researched in women with pre-eclampsia and polycystic ovary syndrome [180], however the overlap with respect to delirium and its associated conditions has not been well addressed. Of the 71 delirium studies, only five studies sought to determine the association with the participants’ common primary condition in their analysis. Tomasi et al. (2017) found that biomarkers differed between patients in the three groups in those with sepsis alone and those who developed sepsis-associated encephalopathy, or delirium,



**Fig. 3** Quality assessment graph of the assay procedures: review author's judgements about each assay domain of the REMARK checklist, presented as percentages across studies

suggesting different mechanisms of sepsis-associated encephalopathy, delirium in people with sepsis, and sepsis itself. Likewise, Pfister et al. (2008) found differences in CRP, s100 calcium binding protein B (s100B) and cortisol in patients with sepsis-associated delirium, compared to non-sepsis associated delirium. In two studies, delirium in stroke was examined [25, 92] but these studies did not identify differences in cortisol [92] or TNF-  $\alpha$ , IL- 1 $\beta$ , IL-18, Brain-derived neurotrophic factor (BDNF) and Neuron specific enolase (NSE) [25] between patients who developed delirium after stroke compared to those who did not develop delirium. Moreover, Sun et al. (2016) attempted to explore the overlap of biomarkers in delirium and dementia in patients with cancer, however, no multivariate analysis was undertaken, therefore results of this study are inconclusive.

Although the aim of this systematic review was to explore the overlap of biomarkers in delirium and advanced cancer syndromes, the findings highlighted a bigger problem in the methodology of delirium biomarker research. The quality assessment in this systematic review found that many of the included studies were of poor methodological quality, inadequately reported, or were influenced by potential confounding factors. A potential barrier to the complete understanding of delirium pathophysiology is the lack of guidelines for conducting and reporting delirium biomarker studies. Results from this review indicate that the absence of such guidelines has likely impeded the quality of individual studies and the overall quality of this critical field of delirium research. Reporting guidelines for delirium biomarker research are an essential step to improving methodological and reporting rigor, and will increase the potential for synthesis of future studies through meta-analyses.

Several studies have previously been performed to determine biomarkers associated with delirium, however potential confounding factors could be the underlying precipitants of delirium; ie risk factors (sepsis), or

underlying conditions present (for example cancer or dementia). The top five most commonly studies biomarkers in this review were inflammatory biomarkers, namely, CRP, IL-6, TNF-  $\alpha$ , IL-10 and IL-8. The challenge with inflammatory markers is that they are non-specific and the inflammatory pathways are similar to those implicated in other conditions such as sepsis and depression [181, 182]. Likewise, of the six delirium studies where there was concomitant cancer, it is very difficult to determine whether those biomarkers found were related to the cancer or the delirium itself, considering alterations in inflammatory pathways are implicated in both. Therefore, future delirium biomarker studies need to be prospectively evaluated and take into account and assess robustly other active co-morbidities such as cancer that could plausibly impact on the pathophysiological and/or biological findings. Similarly, future cancer biomarker studies must also take into account how delirium may clinically or biologically confound biomarker studies in cancer, considering the high prevalence of delirium in this population. Of the six delirium studies with cancer, three did not report the type of cancer, and of the remaining three studies, none were primary brain tumours or brain metastases. Understanding the spread of brain cancer is important in delirium studies, and is an important consideration for future delirium biomarker studies.

Majority of the studies in this review (n=98; 65%) undertook a multivariate analysis, taking into account confounding variables. Where studies only undertook univariate analysis, it is uncertain whether any observed changes in biomarkers were related to the delirium itself, or whether these changes may have been lost when adjusted for confounding factors (such as prior cognitive impairment) in a multivariate analysis. Furthermore, there is likely to be a higher proportion of participants with both delirium and cancer in both groups of studies for which this clinical information was not assessed or that were not reported. Key methodological issues which need to be addressed in future delirium studies include

adjusting for confounders such as age, gender, concurrent medication, comorbidities, prior cognitive impairment, frailty and other neurological conditions. These clinical covariates must also be clearly defined and justified. Assay procedures ought to be reported in detail, including a detailed protocol of the reagents/kits used, repeatability assessments, methods of preservation and storage, assay validity, sensitivity limits of the assay and a scoring and reporting protocol. The timing of the assay is crucial in delirium studies, and the fluctuating pathophysiological processes occurring during delirium, after delirium resolution, and in those who have not yet developed delirium, must be taken into consideration, and be separated in future studies. More standardised and detailed methods of delirium biomarker studies is a crucial step in carrying out future subgroup analyses within this cohort and improving the overall understanding of delirium pathophysiology.

Limitations are that only English language and published studies were included. It is possible that articles were missed; however, two reviewers independently screened all citations derived from a search of six relevant and diverse databases, and all reference lists of included articles were also searched. Another limitation of our study is the lack of a risk of bias tool for biomarker studies, therefore we used an adaptation of tumor marker reporting guidelines, the REMARK checklist [23]. Lastly, the heterogeneity of the data precluded the conduct of a meta-analysis, and precluded any firm conclusions about the biomarkers in delirium and cancer, thus, limiting the rigor of this review. Strengths of this review however, were that we undertook a systematic approach adhering to the PRISMA [15] and an extensive quality assessment of the included studies was undertaken.

## Conclusion

This review found that there is large overlap in the biomarkers in delirium and in advanced cancer-related syndromes, although because of the heterogeneity of the studies firm conclusions about the true overlap of delirium and advanced cancer syndrome biomarkers was not possible. More robust conduct and reporting of delirium biomarker studies will help to better understand the pathophysiology of delirium in the context of co-existing pathophysiology. An improved understanding of the clinical and biological associations of delirium and advanced cancer syndromes in future prospective studies will provide and inform the directions of research into delirium in people with advanced cancer.

## Supplementary information

**Supplementary information** accompanies this paper at <https://doi.org/10.1186/s12888-020-02584-2>.

**Additional file 1:** MEDLINE search strategies MEDLINE search strategies for delirium and cancer studies.

**Additional file 2:** Participant characteristics- delirium studies Characteristics of participants in the included delirium studies.

**Additional file 3:** Participant characteristics- cancer studies Characteristics of participants in the included cancer studies.

**Additional file 4:** Quality assessment of included delirium studies using the REMARK checklist The quality assessment for all included delirium studies.

**Additional file 5:** Quality assessment of included cancer studies using the REMARK checklist The quality assessment for all included cancer studies.

**Additional file 6:** PRISMA checklist.

## Abbreviations

BDNF: Brain-derived neurotrophic factor; CRP: C-reactive protein; CSF: Cerebrospinal fluid; ELISA: Enzyme-linked immunosorbent assay; IL- : Interleukin; NSE: Neuron specific enolase; S100B: S100B calcium binding protein B; TNF: Tumor necrosis factor

## Acknowledgements

Not applicable.

## Ethics approval and consent to participant

Not applicable.

## Authors' contributions

IAD undertook the literature search, identified potential articles, extracted data, interpreted results, performed a quality assessment, drafted and revised all versions of the manuscript. MA and AH contributed to study selection and screening, interpreting results, revised manuscript drafts and supervised the study. All authors (IAD, AH, MA and GC) contributed to the interpretation of results, manuscript preparation and read and approved the final manuscript.

## Funding

None.

## Availability of data and materials

All data generated or analysed in this systematic review are included within this published article and its additional files.

## Consent for publication

Not applicable.

## Competing interests

The authors declare that they have no competing interests.

## Author details

<sup>1</sup>University of Technology Sydney, Faculty of Health, IMPACCT -Improving Palliative, Aged and Chronic Care through Clinical Research and Translation, Sydney, NSW, Australia. <sup>2</sup>Prince of Wales Clinical School, University of New South Wales, Sydney, NSW, Australia. <sup>3</sup>Department of Geriatric Medicine, Prince of Wales Hospital, Sydney, NSW, Australia. <sup>4</sup>South West Sydney Clinical School, University of New South Wales, Liverpool, New South Wales, Australia. <sup>5</sup>Clinical Trials, Ingham Institute of Applied Medical Research, Liverpool, New South Wales, Australia.

Received: 31 July 2019 Accepted: 5 April 2020

Published online: 22 April 2020

## References

- Hosie A, Davidson P, Agar M, Sanderson C, Phillips J. Delirium prevalence, incidence, and implications for screening in specialist palliative care inpatient settings: a systematic review. *Palliat Med*. 2013;27(6):486–98.
- Davis DH, Skelly DT, Murray C, Hennessy E, Bowen J, Norton S, et al. Worsening cognitive impairment and neurodegenerative pathology progressively increase risk for delirium. *Am J Geriatr Psychiatry*. 2015;23(4):403–15.

3. Inouye SK, Westendorp RG, Saczynski JS. Delirium in elderly people. *Lancet*. 2014;383(9920):911–22.
4. American Psychiatric Association. Diagnostic and statistical manual of mental disorders, fifth edition (DSM-5). Arlington: American Psychiatric Publisher; 2013.
5. National Clinical Guideline Centre for Acute and Chronic Conditions. Delirium: diagnosis, prevention and management: NICE clinical guideline 103; 2010. Available from: <https://www.nice.org.uk/guidance/cg103>.
6. Neefjes EC, van der Vorst MJ, Verdegaal BA, Beekman AT, Berkhof J, Verheul HM. Identification of patients with cancer with a high risk to develop delirium. *Cancer Med*. 2017;6(8):1861–70.
7. Uchida M, Okuyama T, Ito Y, Nakaguchi T, Miyazaki M, Sakamoto M, et al. Prevalence, course and factors associated with delirium in elderly patients with advanced cancer: a longitudinal observational study. *Jpn J Clin Oncol*. 2015;45(10):934–40.
8. Grandahl MG, Nielsen SE, Koerner EA, Schultz HH, Arnfred SM. Prevalence of delirium among patients at a cancer ward: clinical risk factors and prediction by bedside cognitive tests. *Nordic J Psychiatry*. 2016;70(6):413–7.
9. Bush SH, Lawlor PG, Ryan K, Centeno C, Lucchesi M, Kanji S, et al. Delirium in adult cancer patients: ESMO clinical practice guidelines. *Ann Oncol*. 2018; 29(Supplement\_4):iv143–iv65.
10. Maldonado JR. Delirium pathophysiology: an updated hypothesis of the etiology of acute brain failure. *Int J Geriatr Psychiatry*. 2017;33(11):1428–57.
11. Berr C. Cognitive impairment and oxidative stress in the elderly: results of epidemiological studies. *Biofactors*. 2000;13(1-4):205–9.
12. Haggstrom L, Nelson J, Wegner E, Caplan G. 2-18F-fluoro-2-deoxyglucose positron emission tomography in delirium. *J Cereb Blood Flow Metab*. 2017; 37(11):3556–67.
13. Caplan GA, Kvelde T, Lai C, Yap SL, Lin C, Hill MA. Cerebrospinal fluid in long-lasting delirium compared with Alzheimer's dementia. *J Gerontol Series A-Med Sci*. 2010;65(10):1130–6.
14. National Cancer Institute. NCI Dictionary of Cancer Terms [Available from: <https://www.cancer.gov/publications/dictionaries/cancer-terms/?CdrID=45618>].
15. Liberati A, Altman DG, Tetzlaff J, Mulrow G, Gøtzsche PC, Ioannidis JP, et al. The PRISMA statement for reporting systematic reviews and meta-analyses of studies that evaluate health care interventions: explanation and elaboration. *PLoS Med*. 2009;6(7):e1000100.
16. Blum D, Omlin A, Fearon K, Baracos V, Radbruch L, Kaasa S, et al. Evolving classification systems for cancer cachexia: ready for clinical practice? *Support Care Cancer*. 2010;18(3):273–9.
17. Bower JE. Cancer-related fatigue—mechanisms, risk factors, and treatments. *Clin Oncol*. 2014;11(10):597.
18. International Association for the Study of Pain. IASP terminology Washington, USA; 2017. Available from: <https://www.iasp-pain.org/Education/Content.aspx?ItemNumber=1698&navItemNumber=576-Pain>.
19. Bray V, Dhillon H, Vardy J. Cancer-related cognitive impairment in adult cancer survivors: a review of the literature cancer forum; 2017. Available from: <https://cancerforum.org.au/forum/2017/march/cancer-related-cognitive-impairment-in-adult-cancer-survivors-a-review-of-the-literature/>.
20. Dantzer R. Cytokine-induced sickness behaviour: a neuroimmune response to activation of innate immunity. *Eur J Pharmacol*. 2004;500(1-3):399–411.
21. Dantzer R. Cytokine-induced sickness behavior: where do we stand? *Brain Behav Immun*. 2001;15(1):7–24.
22. National Cancer Institute. Understanding cancer prognosis; 2018. Available from: <https://www.cancer.gov/about-cancer/diagnosis-staging/prognosis>.
23. Altman DG, McShane LM, Sauerbrei W, Taube SE. Reporting recommendations for tumor marker prognostic studies (REMARK): explanation and elaboration. *BMC Med*. 2012;10(1):51.
24. Egberts A, Mattace-Raso F. Increased neutrophil-lymphocyte ratio in delirium: a pilot study. *Clin Interv Aging*. 2017;12:1115.
25. Kozak HH, Uguz F, Kilinc I, Uca AU, Serhat Tokgoz O, Akpinar Z, et al. Delirium in patients with acute ischemic stroke admitted to the non-intensive stroke unit: incidence and association between clinical features and inflammatory markers. *Neurol Neurochir Pol*. 2017;51(1):38–44.
26. Tomasi CD, Vuolo F, Generoso J, Soares M, Barichello T, Quevedo J, et al. Biomarkers of delirium in a low-risk community-acquired pneumonia-induced sepsis. *Mol Neurobiol*. 2017;54(1):722–6.
27. Vasunilashorn SM, Dillon ST, Inouye SK, Ngo LH, Fong TG, Jones RN, et al. High C-reactive protein predicts delirium incidence, duration, and feature severity after major noncardiac surgery. *J Am Geriatr Soc*. 2017;65(8):e109.
28. Dillon ST, Vasunilashorn SM, Ngo L, Otu HH, Inouye SK, Jones RN, et al. Higher C-reactive protein levels predict postoperative delirium in older patients undergoing major elective surgery: a longitudinal nested case-control study. *Biol Psychiatry*. 2017;81(2):145–53.
29. Guo Y, Jia P, Zhang J, Wang X, Jiang H, Jiang W. Prevalence and risk factors of postoperative delirium in elderly hip fracture patients. *J Int Med Res*. 2016;44(2):317–27.
30. Karlicic IS, Stasevic M, Jankovic S, Dejanovic SD, Milovanovic S. Markers of inflammation as risk predictors of lethal outcome in patients diagnosed with delirium. *Vojnosanit Pregl*. 2016;73(9):838–43.
31. Neerland BE, Hall RJ, Seljeflot I, Frihagen F, MacLulich AMJ, Ræder J, et al. Associations between delirium and preoperative cerebrospinal fluid C-reactive protein, Interleukin-6, and Interleukin-6 receptor in individuals with acute hip fracture. *J Am Geriatr Soc*. 2016;64(7):1456–63.
32. Shen H, Shao Y, Chen J, Guo J. Insulin-like growth factor-1, a potential predictive biomarker for postoperative delirium among elderly patients with open abdominal surgery. *Curr Pharm Design*. 2016;22(38):5879–83.
33. Sun L, Jia P, Zhang J, Zhang X, Zhang Y, Jiang H, et al. Production of inflammatory cytokines, cortisol, and Abeta1-40 in elderly oral cancer patients with postoperative delirium. *Neuropsychiatr Dis Treat*. 2016;12:2789–95.
34. Yen TE, Allen JC, Rivelli SK, Patterson SC, Metcalf MR, Flink BJ, et al. Association between serum IGF-1 levels and postoperative delirium in elderly subjects undergoing elective knee arthroplasty. *Sci Rep*. 2016;6:20736.
35. Avila-Funes JA, Ledesma-Heyer JP, Navarrete-Reyes AP, Chavira-Ramirez R, Boeck-Quirasco L, Aguilar-Navarro S. Association between high serum estradiol levels and delirium among hospitalized elderly women. *Rev Investig Clin*. 2015;67(1):20–4.
36. Brum C, Stertz L, Borba E, Rumi D, Kapczynski F, Camozzato A. Association of serum brain-derived neurotrophic factor (BDNF) and tumor necrosis factor-alpha (TNF-alpha) with diagnosis of delirium in oncology inpatients. *Rev Bras Psiquiatr*. 2015;37(3):197–202.
37. Egberts A, Wijnveld EH, Fekkes D, van der Ploeg MA, Ziere G, Hooijkaas H, et al. Neopterin: a potential biomarker for delirium in elderly patients. *Dement Geriatr Cogn Disord*. 2015;39(1-2):116–24.
38. Foroughan M, Delbari A, Said SE, AkbariKamrani AA, Rashedi V, Zandi T. Risk factors and clinical aspects of delirium in elderly hospitalized patients in Iran. *Aging Clin Exp Res*. 2016;28(2):313–9.
39. Skrede K, Wyller TB, Watne LO, Seljeflot I, Juliebo V. Is there a role for monocyte chemoattractant protein-1 in delirium? Novel observations in elderly hip fracture patients. *BMC Res Notes*. 2015;8:186.
40. Vasunilashorn SM, Ngo L, Inouye SK, Libermann TA, Jones RN, Alsop DC, et al. Cytokines and postoperative delirium in older patients undergoing major elective surgery. *J Gerontol Series A-Med Sci*. 2015;70(10):1289–95.
41. Alexander SA, Ren D, Gunn SR, Kochanek PM, Tate J, Ikonovic M, et al. Interleukin 6 and apolipoprotein E as predictors of acute brain dysfunction and survival in critical care patients. *Am J Crit Care*. 2014;23(1):49–57.
42. Baranyi A, Rothenhausler HB. The impact of soluble interleukin-2 receptor as a biomarker of delirium. [erratum appears in psychosomatics. 2014 Jul-Aug; 55(44):418-9]. *Psychosomatics*. 2014;55(1):51–60.
43. Cape E, Hall RJ, van Munster BC, de Vries A, Howie SE, Pearson A, et al. Cerebrospinal fluid markers of neuroinflammation in delirium: a role for interleukin-1beta in delirium after hip fracture. *J Psychosom Res*. 2014;77(3): 219–25.
44. Capri M, Yani SL, Chattat R, Fortuna D, Buccì L, Lanzarini C, et al. Preoperative, high IL-6 blood level is a risk factor of postoperative delirium onset in old patients. *Front Endocrinol*. 2014;5:173 (SEP) (no pagination).
45. Chen XW, Shi JW, Yang PS, Wu ZQ. Preoperative plasma leptin levels predict delirium in elderly patients after hip fracture surgery. *Peptides*. 2014;57:31–5.
46. Hatta K, Kishi Y, Takeuchi T, Wada K, Odawara T, Usui C, et al. The predictive value of a change in natural killer cell activity for delirium. *Prog Neuro-Psychopharmacol Biol Psychiatry*. 2014;48:26–31.
47. Kazmierski J, Banys A, Latek J, Bourke J, Jaszewski R, Sobow T, et al. Mild cognitive impairment with associated inflammatory and cortisol alterations as independent risk factor for postoperative delirium. *Dement Geriatr Cogn Disord*. 2014;38(1-2):65–78.
48. Ritchie CW, Newman TH, Leurent B, Sampson EL. The association between C-reactive protein and delirium in 710 acute elderly hospital admissions. *Int Psychogeriatr*. 2014;26(5):717–24.
49. Ritter C, Tomasi CD, Dal-Pizzol F, Pinto BB, Dyson A, de Miranda AS, et al. Inflammation biomarkers and delirium in critically ill patients. *Crit Care*. 2014;18(3):R106.

50. Zhang Z, Pan L, Deng H, Ni H, Xu X. Prediction of delirium in critically ill patients with elevated C-reactive protein. *J Crit Care*. 2014;29(1):88–92.
51. Cerejeira J, Batista P, Nogueira V, Vaz-Serra A, Mukaetova-Ladinska EB. The stress response to surgery and postoperative delirium: evidence of hypothalamic-pituitary-adrenal axis hyperresponsiveness and decreased suppression of the GH/IGF-1 Axis. *J Geriatr Psychiatry Neurol*. 2013;26(3):185–94.
52. Colkesen Y, Giray S, Ozenli Y, Sezgin N, Coskun I. Relation of serum cortisol to delirium occurring after acute coronary syndromes. *Am J Emerg Med*. 2013;31(1):161–5.
53. Kazmierski J, Banys A, Latek J, Bourke J, Jaszewski R. Cortisol levels and neuropsychiatric diagnosis as markers of postoperative delirium: a prospective cohort study. *Crit Care*. 2013;17(2):R38.
54. Kazmierski J, Banys A, Latek J, Bourke J, Jaszewski R. Raised IL-2 and TNF-alpha concentrations are associated with postoperative delirium in patients undergoing coronary-artery bypass graft surgery. *Int Psychogeriatr*. 2014;26(5):845–55.
55. Liu P, Li YW, Wang XS, Zou X, Zhang DZ, Wang DX, et al. High serum interleukin-6 level is associated with increased risk of delirium in elderly patients after noncardiac surgery: a prospective cohort study. *Chin Med J*. 2013;126(19):3621–7.
56. Plaschke K, Hautz S, Jansen C, Bruckner T, Schramm C, Karck M, et al. The influence of preoperative serum anticholinergic activity and other risk factors for the development of postoperative cognitive dysfunction after cardiac surgery. *J Thorac Cardiovasc Surg*. 2013;145(3):805–11.
57. Skrobik Y, Leger C, Cossette M, Michaud V, Turgeon J. Factors predisposing to coma and delirium: fentanyl and midazolam exposure; CYP3A5, ABCB1, and ABCG2 genetic polymorphisms; and inflammatory factors. *Crit Care Med*. 2013;41(4):999–1008.
58. Westhoff D, Witlox J, Koenderman L, Kalisvaart KJ, de Jonghe JF, van Stijn MF, et al. Preoperative cerebrospinal fluid cytokine levels and the risk of postoperative delirium in elderly hip fracture patients. *J Neuroinflammation*. 2013;10:122.
59. Bakker RC, Osse RJ, Tulen JH, Kappetein AP, Bogers AJ. Preoperative and operative predictors of delirium after cardiac surgery in elderly patients. *Eur J Cardiothorac Surg*. 2012;41(3):544–9.
60. Baranyi A, Rothenhauser HB. The impact of intra- and postoperative albumin levels as a biomarker of delirium after cardiopulmonary bypass: results of an exploratory study. *Psychiatry Res*. 2012;200(2-3):957–63.
61. Cerejeira J, Nogueira V, Luis P, Vaz-Serra A, Mukaetova-Ladinska EB. The cholinergic system and inflammation: common pathways in delirium pathophysiology. *J Am Geriatr Soc*. 2012;60(4):669–75.
62. Girard TD, Ware LB, Bernard GR, Pandharipande PP, Thompson JL, Shintani AK, et al. Associations of markers of inflammation and coagulation with delirium during critical illness. *Intensive Care Med*. 2012;38(12):1965–73.
63. Osse RJ, Fekkes D, Tulen JH, Wierdsma AI, Bogers AJ, van der Mast RC, et al. High preoperative plasma norepinephrine predicts delirium after cardiac surgery in older adults. *J Am Geriatr Soc*. 2012;60(4):661–8.
64. Bisschop PH, de Rooij SE, Zwinderman AH, van Oosten HE, van Munster BC. Cortisol, insulin, and glucose and the risk of delirium in older adults with hip fracture. *J Am Geriatr Soc*. 2011;59(9):1692–6.
65. Holmes C, Cunningham C, Zotova E, Culliford D, Perry VH. Proinflammatory cytokines, sickness behavior, and Alzheimer disease. *Neurology*. 2011;77(3):212–8.
66. Lee HJ, Hwang DS, Wang SK, Chee IS, Baeg S, Kim JL. Early assessment of delirium in elderly patients after hip surgery. *Psychiatry Investig*. 2011;8(4):340–7.
67. McGrane S, Girard TD, Thompson JL, Shintani AK, Woodworth A, Ely EW, et al. Procalcitonin and C-reactive protein levels at admission as predictors of duration of acute brain dysfunction in critically ill patients. *Crit Care*. 2011;15(2):R78.
68. Morandi A, Gunther ML, Pandharipande PP, Jackson JC, Thompson JL, Shintani AK, et al. Insulin-like growth factor-1 and delirium in critically ill mechanically ventilated patients: a preliminary investigation. *Int Psychogeriatr*. 2011;23(7):1175–81.
69. van den Boogaard M, Kox M, Quinn K, van Achterberg T, van der Hoeven J, Schoonhoven L, et al. Biomarkers associated with delirium in critically ill patients and their relation with long-term subjective cognitive dysfunction; indications for different pathways governing delirium in inflamed and non-inflamed patients. *Crit Care*. 2011;15:R297.
70. van den Boogaard M, van Swelm RPL, Russel FGM, Heemskerk S, van der Hoeven JG, Masereeuw R, et al. Urinary protein profiling in hyperactive delirium and non-delirium cardiac surgery ICU patients. *Proteome Sci*. 2011;9:13 no pagination.
71. Burkhart C, Dell-Kuster S, Gamberini M, Moeckli A, Grapow M, Filipovic M, et al. Modifiable and nonmodifiable risk factors for postoperative delirium after cardiac surgery with cardiopulmonary bypass. *J Cardiothorac Vasc Anesth*. 2010;24(4):555–9 Available from: <http://onlinelibrary.wiley.com/doi/10.1177/0885066610381538/frame.html>.
72. Mu DL, Wang DX, Li LH, Shan GJ, Li J, Yu QJ, et al. High serum cortisol level is associated with increased risk of delirium after coronary artery bypass graft surgery: a prospective cohort study. *Crit Care*. 2010;14:6.
73. Pearson A, de Vries A, Middleton S, Gillies F, White T, Armstrong I, et al. Cerebrospinal fluid cortisol levels are higher in patients with delirium versus controls. *BMC Res Notes*. 2010;3(1):33.
74. Plaschke K, Fichtenkamm P, Schramm C, Hautz S, Martin E, Verch M, et al. Early postoperative delirium after open-heart cardiac surgery is associated with decreased bispectral EEG and increased cortisol and interleukin-6. *Intensive Care Med*. 2010;36(12):2081–9.
75. Tsuruta R, Nakahara T, Miyauchi T, Kutsuna S, Ogino Y, Yamamoto T, et al. Prevalence and associated factors for delirium in critically ill patients at a Japanese intensive care unit. *Gen Hosp Psychiatry*. 2010;32(6):607–11.
76. van Munster BC, Bisschop PH, Zwinderman AH, Korevaar JC, Endert E, Wiersinga WJ, et al. Cortisol, interleukins and S100B in delirium in the elderly. *Brain Cogn*. 2010;74(1):18–23.
77. Adamis D, Lunn M, Martin FC, Treloar A, Gregson N, Hamilton G, et al. Cytokines and IGF-I in delirious and non-delirious acutely ill older medical inpatients. *Age Ageing*. 2009;38(3):326–251.
78. van Munster BC, Korse CM, de Rooij SE, Bonfrer JM, Zwinderman AH, Korevaar JC. Markers of cerebral damage during delirium in elderly patients with hip fracture. *BMC Neurol*. 2009;9:21.
79. Lemstra AW, Kalisvaart KJ, Vreeswijk R, van Gool WA, Eikelenboom P. Pre-operative inflammatory markers and the risk of postoperative delirium in elderly patients. *Int J Geriatr Psychiatry*. 2008;23(9):943–8.
80. Pfister D, Siegemund M, Dell-Kuster S, Smielewski P, Ruegg S, Strebel SP, et al. Cerebral perfusion in sepsis-associated delirium. *Crit Care*. 2008;12(3):R63.
81. Rudolph JL, Ramlawi B, Kuchel GA, McElhanev JE, Xie D, Sellke FW, et al. Chemokines are associated with delirium after cardiac surgery. *J Gerontol Series A-Med Sci*. 2008;63(2):184–9.
82. Van Munster BC, Korevaar JC, Zwinderman AH, Levi M, Wiersinga WJ, De Rooij SE. Time-course of cytokines during delirium in elderly patients with hip fractures. *J Am Geriatr Soc*. 2008;56(9):1704–9.
83. Adamis D, Treloar A, Martin FC, Gregson N, Hamilton G, Macdonald AJD. APOE and cytokines as biological markers for recovery of prevalent delirium in elderly medical inpatients. *Int J Geriatr Psychiatry*. 2007;22(7):688–94.
84. de Rooij SE, van Munster BC, Korevaar JC, Levi M. Cytokines and acute phase response in delirium. *J Psychosom Res*. 2007;62(5):521–5.
85. Plaschke K, Hill H, Engelhardt R, Thomas C, von Haken R, Scholz M, et al. EEG changes and serum anticholinergic activity measured in patients with delirium in the intensive care unit. *Anaesthesia*. 2007;62(12):1217–23.
86. White S, Calver BL, Newsway V, Wade R, Patel S, Bayer A, et al. Enzymes of drug metabolism during delirium. *Age Ageing*. 2005;34(6):603–8.
87. Wilson K, Broadhurst C, Diver M, Jackson M, Mottram P. Plasma insulin growth factor-1 and incident delirium in older people. *Int J Geriatr Psychiatry*. 2005;20(2):154–9.
88. Beloosesky Y, Grinblat J, Pirotsky A, Weiss A, Hendel D. Different C-reactive protein kinetics in post-operative hip-fractured geriatric patients with and without complications. *Gerontology*. 2004;50(4):216–22.
89. Robertsson B, Blennow K, Brane G, Edman A, Karlsson I, Wallin A, et al. Hyperactivity in the hypothalamic-pituitary-adrenal axis in demented patients with delirium. *Int Clin Psychopharmacol*. 2001;16(1):39–47.
90. van der Mast RC, van den Broek WW, Fekkes D, Pepplinkhuizen L, Habbema JD. Is delirium after cardiac surgery related to plasma amino acids and physical condition? *J Neuropsychiatr Clin Neurosci*. 2000;12(1):57–63.
91. van der Mast RC, van den Broek WW, Fekkes D, Pepplinkhuizen L, Habbema JDF. Incidence of and preoperative predictors for delirium after cardiac surgery. *J Psychosom Res*. 1999;46(5):479–83.
92. Gustafson Y, Olsson T, Asplund K, Hägg E. Acute confusional state (delirium) soon after stroke is associated with hypercortisolism. *Cerebrovasc Dis*. 1993;3(1):33–8.
93. McIntosh TK, Bush HL, Yeston NS. Beta-endorphin, cortisol and postoperative delirium: a preliminary report. *Psychoneuroendocrinology*. 1985;10(3):303–13.
94. Amano K, Maeda I, Morita T, Baba M, Miura T, Hama T, et al. C-reactive protein, symptoms and activity of daily living in patients with advanced cancer receiving palliative care. *J Cachexia Sarcopenia Muscle*. 2017;8(3):457.

95. Demiray G, DeGirmencioGlu S, Ugurlu E, Yaren A. Effects of serum leptin and resistin levels on cancer cachexia in patients with advanced-stage non-small cell lung cancer. *Clin Med Insights*. 2017;11:1 (no pagination) (1179554917690144).
96. Fogelman DR, Morris J, Xiao L, Hassan M, Vadhan S, Overman M, et al. A predictive model of inflammatory markers and patient-reported symptoms for cachexia in newly diagnosed pancreatic cancer patients. *Support Care Cancer*. 2017;25(6):1809–17.
97. Luo Y, Kim HS, Kim M, Lee M, Song YS. Elevated plasma fibrinogen levels and prognosis of epithelial ovarian cancer: a cohort study and meta-analysis. *J Gynecol Oncol*. 2017;28:3.
98. Paulsen O, Laird B, Aass N, Lea T, Fayers P, Kaasa S, et al. The relationship between pro-inflammatory cytokines and pain, appetite and fatigue in patients with advanced cancer. *PLoS One*. 2017;12(5):e0177620 no pagination.
99. Amano K, Maeda I, Morita T, Miura T, Inoue S, Ikenaga M, et al. Clinical implications of C-reactive protein as a prognostic marker in advanced cancer patients in palliative settings. *Eur J Cancer*. 2016;51:S207.
100. Bye A, Wesseltoft-Rao N, Iversen PO, Skjægstad G, Holven KB, Ulven S, et al. Alterations in inflammatory biomarkers and energy intake in cancer cachexia: a prospective study in patients with inoperable pancreatic cancer. *Med Oncol*. 2016;33(6):54.
101. Mitsunaga S, Ikeda M, Shimizu S, Ohno I, Takahashi H, Okuyama H, et al. C-reactive protein level is an indicator of the aggressiveness of advanced pancreatic cancer. *Pancreas*. 2016;45(1):110–6.
102. Morgado PC, Giorlando A, Castro M, Navigante A. Relationship between weight loss and parameters of skeletal muscle function in patients with advanced cancer and fatigue. *Support Care Cancer*. 2016;24(9):3961–6.
103. Rodrigues AR, Truffelli DC, Fonseca F, de Paula LC. Giglio ad. Fatigue in patients with advanced terminal cancer correlates with inflammation, poor quality of life and sleep, and anxiety/depression. *Am J Hosp Palliat Med*. 2016;33(10):942–7.
104. Srdic D, Plestina S, Sverko-Peternac A, Nikolac N, Simundic AM, Samarzija M. Cancer cachexia, sarcopenia and biochemical markers in patients with advanced non-small cell lung cancer-chemotherapy toxicity and prognostic value. *Support Care Cancer*. 2016;24(11):4495–502.
105. Wu Y, Li C, Zhao J, Yang L, Liu F, Zheng H, et al. Neutrophil-to-lymphocyte and platelet-to-lymphocyte ratios predict chemotherapy outcomes and prognosis in patients with colorectal cancer and synchronous liver metastasis. *World J Surg Oncol*. 2016;14(1):289.
106. Bilir C, Engin H, Can M, Temi YB, Demirtas D. The prognostic role of inflammation and hormones in patients with metastatic cancer with cachexia. *Med Oncol*. 2015;32(3):56.
107. Miura T, Mitsunaga S, Ikeda M, Shimizu S, Ohno I, Takahashi H, et al. Characterization of patients with advanced pancreatic cancer and high serum interleukin-6 levels. *Pancreas*. 2015;44(5):756–63.
108. Miura T, Matsumoto Y, Hama T, Amano K, Tei Y, Kikuchi A, et al. Glasgow prognostic score predicts prognosis for cancer patients in palliative settings: a subanalysis of the Japan–prognostic assessment tools validation (J-ProVal) study. *Support Care Cancer*. 2015;23(11):3149–56.
109. Barrera L, Montes-Servin E, Barrera A, Ramirez-Tirado L, Salinas-Parra F, Banales-Mendez J, et al. Cytokine profile determined by data-mining analysis set into clusters of non-small-cell lung cancer patients according to prognosis. *Ann Oncol*. 2014;26(2):428–35.
110. Blakely AM, Heffernan DS, McPhillips J, Cioffi WG, Miner TJ. Elevated C-reactive protein as a predictor of patient outcomes following palliative surgery. *J Surg Oncol*. 2014;110(6):651–5.
111. Fujiwara Y, Kobayashi T, Chayahara N, Imamura Y, Toyoda M, Kiyota N, et al. Metabolomics evaluation of serum markers for cachexia and their intra-day variation in patients with advanced pancreatic cancer. *PLoS One*. 2014;9(11):e113259.
112. Lindenmann J, Neubock N, Smolle J, Maier A, Smolle-Juttner FM. The influence of elevated levels of C-reactive protein and hypoalbuminaemia on survival in patients with advanced inoperable oesophageal cancer undergoing palliative treatment. *Eur Surg*. 2014;46:S57.
113. Mondello P, Lacquaniti A, Mondello S, Bolognani D, Pitini V, Aloisi C, et al. Emerging markers of cachexia predict survival in cancer patients. *BMC Cancer*. 2014;14(1):828.
114. Moriwaki T, Ishige K, Araki M, Yoshida S, Nishi M, Sato M, et al. Glasgow prognostic score predicts poor prognosis among advanced biliary tract cancer patients with good performance status. *Med Oncol*. 2014;31(11):287.
115. Szkandera J, Stotz M, Absenger G, Stojakovic T, Samonigg H, Kornprat P, et al. Validation of C-reactive protein levels as a prognostic indicator for survival in a large cohort of pancreatic cancer patients. *Br J Cancer*. 2014;110(1):183.
116. Zhang SY, Zeng D, Peng YH, Yang YX, Zhuang XW, Li ZT, et al. Cancer-related fatigue and chemotherapy-associated adverse effects: correlation with TNF-alpha, IL-1 and 17-hydroxycorticosteroids. *Future Oncol*. 2014;10(9):1619–26.
117. Jafri SH, Shi R, Mills G. Advance lung cancer inflammation index (ALI) at diagnosis is a prognostic marker in patients with metastatic non-small cell lung cancer (NSCLC): a retrospective review. *BMC Cancer*. 2013;13(1):158.
118. Laird BJ, McMillan DC, Fayers P, Fearon K, Kaasa S, Fallon MT, et al. The systemic inflammatory response and its relationship to pain and other symptoms in advanced cancer. *Oncologist*. 2013;18(9):1050–5.
119. Laird BJ, Kaasa S, McMillan DC, Fallon MT, Hjermstad MJ, Fayers P, et al. Prognostic factors in patients with advanced cancer: a comparison of clinicopathological factors and the development of an inflammation-based prognostic system. *Clin Cancer Res*. 2013;19(19):5456–64.
120. Paiva CE, Paiva BSR. Prevalence, predictors, and prognostic impact of fatigue among Brazilian outpatients with advanced cancers. *Support Care Cancer*. 2013;21(4):1053–60.
121. Suh SY, Choi YS, Yeom CH, Kwak SM, Yoon HM, Kim DG, et al. Interleukin-6 but not tumour necrosis factor-alpha predicts survival in patients with advanced cancer. *Support Care Cancer*. 2013;21(11):3071–7.
122. de Raaf P, Sleijfer S, Lamers C, Jager A, Gratama J, van der Rijt C. The association between inflammation and fatigue dimensions in advanced cancer patients and cancer survivors. *Palliat Med*. 2012;26(4):449–50.
123. Gioulbasanis I, Patrikidou A, Kitikidou K, Papadimitriou K, Vlachostergios PJ, Tsatsanis C, et al. Baseline plasma levels of Interleukin-8 in stage IV non-small-cell lung cancer patients: relationship with nutritional status and prognosis. *Nutr Cancer*. 2012;64(1):41–7.
124. Gulen ST, Karadag F, Karul AB, Kilicarslan N, Ceylan E, Kuman NK, et al. Adipokines and systemic inflammation in weight-losing lung cancer patients. *Lung*. 2012;190(3):327–32.
125. Heitzer E, Sandner-Kiesling A, Schippinger W, Stohscheer I, Osprian I, Bitsche S, et al. IL-7, IL-18, MCP-1, MIP1-beta, and OPG as biomarkers for pain treatment response in patients with cancer. *Pain Physician*. 2012;15(6):499–510.
126. Minton O, Strasser F, Radbruch L, Stone P. Identification of factors associated with fatigue in advanced cancer: a subset analysis of the European palliative care research collaborative computerized symptom assessment data set. *J Pain Symptom Manag*. 2012;43(2):226–35.
127. Partridge M, Fallon M, Bray C, McMillan D, Brown D, Laird B. Prognostication in advanced cancer: a study examining an inflammation-based score. *J Pain Symptom Manag*. 2012;44(2):161–7.
128. Pond GR, Armstrong AJ, Wood BA, Leopold L, Galsky MD, Sonpavde G. Ability of C-reactive protein to complement multiple prognostic classifiers in men with metastatic castration resistant prostate cancer receiving docetaxel-based chemotherapy. *BJU Int*. 2012;110:11b.
129. Wang D-s, Luo H-y, M-z Q, Wang Z-q, D-s Z, Wang F-h, et al. Comparison of the prognostic values of various inflammation based factors in patients with pancreatic cancer. *Med Oncol*. 2012;29(5):3092–100.
130. Aydin Y, Kaplan I, Gundogdu B, Albayrak B, Turkiylmaz A, Eroglu A. Prognostic importance of serum CRP, prealbumin, and transferrin levels in patients with advanced stage esophageal cancer. *Turk Gogus Kalp Damar Cerrahisi Derg*. 2011;19(3):384–90.
131. Dev R, Hui D, Dalal S, Nooruddin ZI, Yennurajalingam S, Del Fabbro E, et al. Association between serum cortisol and testosterone levels, opioid therapy, and symptom distress in patients with advanced cancer. *J Pain Symptom Manag*. 2011;41(4):788–95.
132. Gioulbasanis I, Georgoulis P, Vlachostergios PJ, Baracos V, Ghosh S, Giannousi Z, et al. Mini nutritional assessment (MNA) and biochemical markers of cachexia in metastatic lung cancer patients: interrelations and associations with prognosis. *Lung Cancer*. 2011;74(3):516–20.
133. Hwang J-E, Kim H-N, Kim D-E, Choi H-J, Jung S-H, Shim H-J, et al. Prognostic significance of a systemic inflammatory response in patients receiving first-line palliative chemotherapy for recurrent or metastatic gastric cancer. *BMC Cancer*. 2011;11(1):489.
134. Kwak SM, Choi YS, Yoon HM, Kim DG, Song SH, Lee YJ, et al. The relationship between interleukin-6, tumor necrosis factor- $\alpha$ , and fatigue in terminally ill cancer patients. *Palliat Med*. 2012;26(3):275–82.

135. Lee JS, Kwon OY, Choi HS, Hong HP, Ko YG. Serum C-reactive protein level is a predictive factor for 14-day mortality of patients with advanced cancer who present to the emergency department with acute symptoms. *Acad Emerg Med*. 2011;18(4):440–2.
136. Scheede-Bergdahl C, Watt HL, Trutschnigg B, Kilgour RD, Haggarty A, Lucar E, et al. Is IL-6 the best pro-inflammatory biomarker of clinical outcomes of cancer cachexia? *Clin Nutr*. 2012;31(1):85–8.
137. Vlachostergios P, Gioulbasanis I, Kamposioras K, Georgoulas P, Baracos V, Ghosh S, et al. Baseline insulin-like growth factor-I plasma levels, systemic inflammation, weight loss and clinical outcome in metastatic non-small cell lung cancer patients. *Oncol*. 2011;81(2):113–8.
138. Diakowska D, Krzystek-Korpacka M, Markocka-Maczka K, Diakowski W, Matusiewicz M, Grabowski K. Circulating leptin and inflammatory response in esophageal cancer, esophageal cancer-related cachexia-anorexia syndrome (CAS) and non-malignant CAS of the alimentary tract. *Cytokine*. 2010;51(2):132–7.
139. Meek CL, Wallace AM, Forrest LM, McMillan DC. The relationship between the insulin-like growth factor-1 axis, weight loss, an inflammation-based score and survival in patients with inoperable non-small cell lung cancer. *Clin Nutr*. 2010;29(2):206–9.
140. Ishizuka M, Nagata H, Takagi K, Kubota K. Influence of inflammation-based prognostic score on mortality of patients undergoing chemotherapy for far advanced or recurrent unresectable colorectal cancer. *Ann Surg*. 2009; 250(2):268–72.
141. Karapanagiotou EM, Polyzos A, Dilana KD, Gratsias I, Boura P, Gkiozos I, et al. Increased serum levels of ghrelin at diagnosis mediate body weight loss in non-small cell lung cancer (NSCLC) patients. *Lung Cancer*. 2009;66(3):393–8.
142. Paddison JS, Temel JS, Fricchione GL, Pirl WF. Using the differential from complete blood counts as a biomarker of fatigue in advanced non-small-cell lung cancer: an exploratory analysis. *Palliat Support Care*. 2009;7(2):213–7.
143. Takahashi M, Terashima M, Takagane A, Oyama K, Fujiwara H, Wakabayashi G. Ghrelin and leptin levels in cachectic patients with cancer of the digestive organs. *Int J Clin Oncol*. 2009;14(4):315–20.
144. Inagaki M, Isono M, Okuyama T, Sugawara Y, Akechi T, Akizuki N, et al. Plasma interleukin-6 and fatigue in terminally ill cancer patients. *J Pain Symptom Manag*. 2008;35(2):153–61.
145. Karapanagiotou EM, Tsochatzis EA, Dilana KD, Tourkantonis I, Gratsias I, Syrigos KN. The significance of leptin, adiponectin, and resistin serum levels in non-small cell lung cancer (NSCLC). *Lung Cancer*. 2008;61(3):391–7.
146. Sharma R, Zucknick M, London R, Kacevska M, Liddle C, Clarke SJ. Systemic inflammatory response predicts prognosis in patients with advanced-stage colorectal cancer. *Clin Colorectal Cancer*. 2008;7(5):331–7.
147. Weryńska B, Kosacka M, Golecki M, Jankowska R. Leptin serum levels in cachectic and non-cachectic lung cancer patients. *Adv Respi Med*. 2009; 77(6):500–6.
148. Ravasco P, Monteiro-Grillo I, Camilo M. How relevant are cytokines in colorectal cancer wasting? *Cancer J*. 2007;13(6):392–8.
149. Richey LM, George JR, Couch ME, Kanapkey BK, Yin X, Cannon T, et al. Defining cancer cachexia in head and neck squamous cell carcinoma. *Clin Cancer Res*. 2007;13(22):6561–7.
150. Suh S-Y, Ahn H-Y. A prospective study on C-reactive protein as a prognostic factor for survival time of terminally ill cancer patients. *Support Care Cancer*. 2007;15(6):613.
151. Al Murri A, Bartlett J, Canney P, Doughty J, Wilson C, McMillan D. Evaluation of an inflammation-based prognostic score (GPS) in patients with metastatic breast cancer. *Br J Cancer*. 2006;94(2):227.
152. Kayacan O, Karnak D, Beder S, Güllü E, Tutkak H, Senler FC, et al. Impact of TNF- $\alpha$  and IL-6 levels on development of cachexia in newly diagnosed NSCLC patients. *Am J Clin Oncol*. 2006;29(4):328–35.
153. Ramsey S, Lamb GW, Aitchison M, Graham J, McMillan DC. Evaluation of an inflammation-based prognostic score in patients with metastatic renal cancer. *Cancer*. 2007;109(2):205–12.
154. Di Nisio M, Niers TM, Reitsma PH, Buller HR. Plasma cytokine and P-selectin levels in advanced malignancy: prognostic value and impact of low-molecular weight heparin administration. *Cancer*. 2005;104(10):2275–81.
155. Rich T, Innominato PF, Boerner J, Mormont MC, Iacobelli S, Baron B, et al. Elevated serum cytokines correlated with altered behavior, serum cortisol rhythm, and dampened 24-hour rest-activity patterns in patients with metastatic colorectal cancer. *Clin Cancer Res*. 2005;11(5):1757–64.
156. Bolukbas FF, Kilic H, Bolukbas C, Gumus M, Horoz M, Turhal NS, et al. Serum leptin concentration and advanced gastrointestinal cancers: a case controlled study. *BMC Cancer*. 2004;4(1):29.
157. De Vita F, Romano C, Orditura M, Galizia G, Martinelli E, Lieto E, et al. Interleukin-6 serum level correlates with survival in advanced gastrointestinal cancer patients but is not an independent prognostic indicator. *J Interf Cytokine Res*. 2001;21(1):45–52.
158. Dülger H, Alici S, Şekeroğlu M, Erkog R, Özbek H, Noyan T, et al. Serum levels of leptin and proinflammatory cytokines in patients with gastrointestinal cancer. *Int J Clin Pract*. 2004;58(6):545–9.
159. Elahi MM, McMillan DC, McArdle CS, Angerson WJ, Sattar N. Score based on hypoalbuminemia and elevated C-reactive protein predicts survival in patients with advanced gastrointestinal cancer. *Nutr Cancer*. 2004;48(2):171–3.
160. Jamieson NB, Brown DJ, Wallace AM, McMillan DC. Adiponectin and the systemic inflammatory response in weight-losing patients with non-small cell lung cancer. *Cytokine*. 2004;27(2-3):90–2.
161. Songur N, Kuru B, Kalkan F, Özdilekcan C, Cakmak H, Hizel N. Serum interleukin-6 levels correlate with malnutrition and survival in patients with advanced non-small cell lung cancer. *Tumori*. 2004;90(2):196–200.
162. Scott HR, McMillan DC, Brown DJ, Forrest LM, McArdle CS, Milroy R. A prospective study of the impact of weight loss and the systemic inflammatory response on quality of life in patients with inoperable non-small cell lung cancer. *Lung Cancer*. 2003;40(3):295–9.
163. Alemán MR, Santolaria F, Batista N, Maria J, González-Reimers E, Milena A, et al. Leptin role in advanced lung cancer. A mediator of the acute phase response or a marker of the status of nutrition? *Cytokine*. 2002;19(1):21–6.
164. Orditura M, De Vita F, Catalano G, Infusino S, Lieto E, Martinelli E, et al. Elevated serum levels of interleukin-8 in advanced non-small cell lung cancer patients: relationship with prognosis. *J Interf Cytokine Res*. 2002; 22(11):1129–35.
165. Scott H, McMillan D, Forrest L, Brown D, McArdle C, Milroy R. The systemic inflammatory response, weight loss, performance status and survival in patients with inoperable non-small cell lung cancer. *Br J Cancer*. 2002;87(3):264.
166. Jatoi A, Loprinzi CL, Sloan JA, Klee GG, Windschitl HE. Neuropeptide Y, leptin, and cholecystokinin 8 in patients with advanced cancer and anorexia - a north Central cancer treatment group exploratory investigation. *Cancer*. 2001;92(3):629–33.
167. Mantovani G, Maccio A, Madeddu C, Mura L, Massa E, Mudu M, et al. Serum values of proinflammatory cytokines are inversely correlated with serum leptin levels in patients with advanced stage cancer at different sites. *J Mol Med*. 2001;79(7):406–14.
168. Mantovani G, Maccio A, Mura L, Massa E, Mudu MC, Mulas C, et al. Serum levels of leptin and proinflammatory cytokines in patients with advanced-stage cancer at different sites. *J Mol Med*. 2000;78(10):554–61.
169. Nenova K, Kovatchev D. TNF-A levels in cachectic cancer patients. *Arch Hellenic Med*. 2000;17(6):619–22.
170. O'Gorman P, McMillan DC, McArdle CS. Longitudinal study of weight, appetite, performance status, and inflammation in advanced gastrointestinal cancer. *Nutr Cancer*. 1999;35(2):127–9.
171. Okada S, Okusaka T, Ishii H, Kyogoku A, Yoshimori M, Kajimura N, et al. Elevated serum interleukin-6 levels in patients with pancreatic cancer. *Jpn J Clin Oncol*. 1998;28(1):12–5.
172. Wallace AM, Kelly A, Sattar N, McArdle CS, McMillan DC. Circulating concentrations of “free” leptin in relation to fat mass and appetite in gastrointestinal cancer patients. *Nutr Cancer*. 2002;44(2):157–60.
173. Maltoni M, Marco P, Oriana N, Mauro M, Monica I, Gramazio A, et al. Biological indices predictive of survival in 519 Italian terminally ill cancer patients. *J Pain Symptom Manag*. 1997;13:1.
174. Simons J, Schols A, Campfield L, Wouters E, Saris W. Plasma concentration of total leptin and human lung-cancer-associated cachexia. *Clin Sci*. 1997; 93(3):273–7.
175. Wallace A, Sattar N, McMillan D. Effect of weight loss and the inflammatory response on leptin concentrations in gastrointestinal cancer patients. *Clin Cancer Res*. 1998;4(12):2977–9.
176. Marcantonio ER, Rudolph JL, Cully D, Crosby G, Alsop D, Inouye SK. Serum biomarkers for delirium. *J Gerontol Series A-Med Sci*. 2006;61(12):1281–6.
177. Kellum JA, Kong L, Fink MP, Weissfeld LA, Yealy DM, Pinsky MR, et al. Understanding the inflammatory cytokine response in pneumonia and sepsis: results of the genetic and inflammatory markers of sepsis (GenIMS) study. *Arch Intern Med*. 2007;167(15):1655–63.
178. Saribal D, Hocaoglu-Emre F, Erdogan S, Bahtiyar N, Okur SC, Mert M. Inflammatory cytokines IL-6 and TNF- $\alpha$  in patients with hip fracture. *Osteoporos Int*. 2019;30(5):1025–31.

179. Hennessy E, Gormley S, Lopez-Rodriguez AB, Murray C, Murray C, Cunningham C. Systemic TNF- $\alpha$  produces acute cognitive dysfunction and exaggerated sickness behavior when superimposed upon progressive neurodegeneration. *Brain Behav Immun*. 2017;59:233–44.
180. Khan GH, Galazis N, Docheva N, Layfield R, Atiomo W. Overlap of proteomics biomarkers between women with pre-eclampsia and PCOS: a systematic review and biomarker database integration. *Hum Reprod*. 2014; 30(1):133–48.
181. Strawbridge R, Young AH, Cleare AJ. Biomarkers for depression: recent insights, current challenges and future prospects. *Neuropsychiatr Dis Treat*. 2017;13:1245.
182. Cho S-Y, Choi J-H. Biomarkers of sepsis. *Infect Chemother*. 2014;46(1):1–12.

### **Publisher's Note**

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

**Ready to submit your research? Choose BMC and benefit from:**

- fast, convenient online submission
- thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

**At BMC, research is always in progress.**

Learn more [biomedcentral.com/submissions](https://biomedcentral.com/submissions)

